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RESEARCH ARTICLE



# Karyomorphometric analysis of Fritillaria montana group in Greece

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#### Abstract

*Fritillaria* Linnaeus, 1753 (Liliaceae) is a genus of geophytes, represented in Greece by 29 taxa. Most of the Greek species are endemic to the country and/or threatened. Although their classical cytotaxonomic studies have already been presented, no karyomorphometric analysis has ever been given. In the present study, the cytological results of *Fritillaria montana* Hoppe ex W.D.J. Koch, 1832 group, which includes *F epirotica* Turrill ex Rix, 1975 and *F montana* are statistically evaluated for the first time. Further indices about interchromosomal and intrachromosomal asymmetry are given. A new population of *F. epirotica* is also investigated, while for *F. montana*, a diploid individual was found in a known as triploid population. Paired t-tests and PCoA analysis have been applied to compare the two species.

#### Keywords

Fritillaria epirotica, Fritillaria montana, karyotype analysis, PCoA, endemics, Greek flora, karyograms

## Introduction

The genus *Fritillaria* Linnaeus, 1753 (Liliaceae) comprises approximately 150 taxa of geophytes, found in the temperate zones of the Northern Hemisphere (Kamari and Phitos 2006). Most of them are distributed across Eurasia while about 20 species occur in

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California. Only one species, *F. camschatcensis* (L.) Ker Gawler, 1809 links both groups by its distribution in both North America and eastern Asia (Fay and Chase 2000, Ambrozova et al. 2011).

According to its latest revision (Rix 2001), the genus is divided into eight subgenera, *Fritillaria* Rix, 2001 (including two sections, *Olostyleae* Rix, 2001 and *Fritillaria* Rix, 2001); *Davidii* Rix, 2001; *Liliorhiza* (Kellogg) Bentham & Hooker, 1883; *Japonica* Rix, 2001; *Rhinopetalum* Fischer, 1835; *Petilium* Baker, 1874; the monotypic *Theresia* K. Koch, 1849 and *Korolkowia* Rix, 2001. Although Iran (and more precisely its northern part as well as the neighbouring countries) is relatively poor in species (17 species and 4 subspecies), it is considered to be the centre of *Fritillaria* diversity above species level (Rix 1977), because those taxa belong to four out of five main subgenera (Jafari et al. 2014).

In Greece, the genus is also characterized by high diversity and is represented by a multitude of taxa (24 species and 5 subspecies), all belonging to the subgenus *Fritillaria* (Kamari and Phitos 2006).

Out of the 29 taxa found or described in Greece so far, 18 taxa (14 species and 4 subspecies) are endemic to the country and no less than 17 species and 2 subspecies occur in the Aegean archipelago and the surrounding continental region (Kamari and Phitos 2000). Moreover, Turkey is the richest country concerning the number of *Fritillaria* with 35 species and 6 subspecies, 19 of which are considered endemic (Tekşen and Aytaç 2011, Advay et al. 2015, Özhatay et al. 2015). Eighteen of those species and 4 subspecies are distributed in the Mediterranean, 12 of which are endemic. Taking into consideration the total number of *Fritillaria* taxa as well as the number of the endemic ones, Greece, along with W Turkey (Rix 1984, Özhatay 2000, Tekşen 2012), constitutes a secondary evolutionary center at least for this subgenus, if not for the whole genus. As a result, the Aegean archipelago can be considered as the heart of the secondary biodiversity center for the subgenus *Fritillaria* (Kamari and Phitos 2000).

Among the *Fritillaria* taxa occurring in continental Greece two species constitute the *Fritillaria montana* group (Kamari 1991a): *Fritillaria epirotica* Turrill ex Rix, 1975, which is endemic to NW Greece and *Fritillaria montana* Hoppe ex W.D.J.Koch, 1832, which has a wide distribution in S and SE Europe. Both species of the above group are characterized by their long (2/3 of the tepal length) nectaries, as well as by their obscurely tessellated tepals.

*Fritillaria epirotica* is a very short plant (up to 15 cm) with dark purplish, obscurely tessellated flowers, which almost touch the ground and it grows on ophiolithic substrates, usually at high altitudes (up to 2600 m). On the contrary, *F. montana* is tall (up to 60 cm), characterized by alternate or subopposite linear, slightly canaliculated leaves, with dark purplish distinctly tessellated flowers, and it grows usually on limestone substrate at an altitude up to 1600 m.

*Fritillaria epirotica* is included in the Red Data Book of Rare and Threatened Plants of Greece (Phitos et al. 1995 & 2009), in the IUCN Red List of Threatened Species, Version 2014.2. and also in the Council Directive 92/43/EEC on the conservation of natural habitats and of wild fauna and flora. It is protected by the Presidential Decree

67/81, characterized as Endangered (EN) by IUCN and as Vulnerable (VU) in the Red Data Book of Rare and Threatened Plants of Greece (Kamari 1995, Kamari and Phitos 2009). Fritillaria montana is characterized, according to IUCN Red List of Threatened Species, Version 2014.2., as a Data Deficient (DD) species. In Greece, some of its populations are included in Natura 2000 sites. Despite its wide distribution, the species is Rare (R) in Italy (Peruzzi et al. 2008) and included in the regional Red Lists of Italian threatened species (Conti et al. 1997). As already known, the misapplied nomenclature of the F. montana complicates botanical literature (Lozina-Lozinskaja 1935, Zahariadi 1966, Kamari 1991a, b, Tomovic et al. 2007). Several locations in Italy have recently been further studied and Fritillaria montana populations are getting distinguished, while more biometric details for the species are provided (Peruzzi and Bartolucci 2009, Bartolucci et al. 2009, Mancuso et al. 2012, Peruzzi et al. 2012). An indicative example of the situation is the very low production of fruits during fruiting season in 2008 observed by Mancuso et al. (2012). Moreover, Fritillaria montana is characterized as an Endangered (EN) species listed in the third edition of the Red Book of Ukraine (Chorney et al. 2009), as a Rare (R) one in Bosnia and Herzegovina (Šilić 1996), Vulnerable (VU) in Serbia (IUCN Red List of Threatened Species) and protected at a national level in France. Tomovic et al. (2007) referred that the species was listed as Rare in the Red Data Book of the PR Bulgaria (Velchev 1984 sub F. orientalis Adam), but the latest version does not include it anymore (Petrova and Vladimirov 2009).

Concerning the cytology of the genus, Fritillaria has been studied for many years due to the interest of its large chromosomes and vast genome size (Darlington 1935, 1937, Frankel 1940). Indeed, 1C values (DNA content of the unreplicated haploid chromosome complement) in *Fritillaria* are among the largest reported for all angiosperms (Bennett and Smith 1976, Sharma and Raina 2005). The karyotype is quite stable, asymmetrical and usually diploid, with a basic chromosome number of x = 12. Only a few species are an exception to this, with x = 9 (3 species), x = 11 (2 species) and x = 13 (2 species), but without a special pattern (Darlington 1937, Noda 1964, Li and Shang 1989, Jafari et al. 2014). However, the presence and the morphology of satellited chromosomes vary among the species or even among populations of the same taxon (Runemark 1970, Bentzer et al. 1971, Mehra and Sachdeva 1976, Koul and Wafai 1980, Kamari 1984a, 1991a, 1996, Zaharof 1987, Kamari and Phitos 2006). In addition, secondary constrictions and supernumerary B-chromosomes are observed very often (La Cour 1978b, 1978c, Kamari 1984a, 1991a, 1991b, Zaharof 1987, 1989, Kamari and Phitos 2006). As a result, that type of differentiations is always emphasized and specific chromosome pairs are studied as markers, in order to spot the differences among the generally stable and similar karyotypes (Kamari 1984b, Zaharof 1989, Kamari and Phitos 2000, 2006). Finally, a few triploid karyotypes have been reported with 2n = 3x = 36 (Fedorov 1969, La Cour 1978a, Moore 1982, Marchant and Macfarlane 1980, Zaharof 1987, Peruzzi et al. 2009) or with 2n = 3x = 27 chromosomes (Cesca 1986, Kamari 1991a).

Recently many questions have arisen, regarding the classification and phylogeny of the genus, especially for the species appearing in Greece. Although several molecular phylogenetic studies have been published (Fay and Chase 2000, Rønsted et al. 2005, Turktas et al. 2012, Metin et al. 2013) none of them refer to the total of Greek taxa. Even though classical cytotaxonomic studies of the genus in Greece have already been published (Kamari 1984a, 1984b, 1991a, 1991b, 1996, Zaharof 1987, 1989, Kamari and Phitos 2000, 2006), neither karyomorphometric analysis, nor statistical evaluation of the cytological results, have ever been given so far. In the present study, an attempt for further karyomorphometric analysis of chromosome features has been made, concerning the two members of *Fritillaria montana* group.

#### Material and methods

Living plants of the *Fritillaria montana* and *F. epirotica* populations were collected (Table 1) and cultivated in the Experimental Gardens of the University of Patras and Agricultural University of Athens. Vouchers are deposited in UPA and ACA.

The cytological study is based on the squash technique and the chromosome counts were made from root tip metaphases (Östergren and Heneen 1962, Kamari 1976). The root tips were pretreated in a mixture of 1:1 8-hydroxyquinoline (0,002% w/v):colchicine (0.3 w/v) for 3 hrs (Kamari 1984a) and fixed in 3:1 (v/v) absolute ethanol:glacial acetic acid for 24 hours at 4 °C. Fixed root tips were stored at -20 °C at 75% ethanol.

Before staining, the root tips were hydrolyzed in 1N HCl 60 °C for 15 min and stained in Feulgen for 3 hrs (Darlington and La Cour 1969). Prior to squashing, the stained root tips were put on a slide with a drop of 45% (v/v) acetic acid. The slides were observed with AXIOLAB Zeiss microscope and photos were taken with Canon EOS 600D digital camera.

At least five metaphase plates of each species were analysed and indices were calculated with Microsoft Office Excel 2007, IBM SPSS Statistics version 22 and Past 3.03. Chromosome terminology follows Levan et al. (1965), Stebbins (1971) and Kamari (1976), taking into consideration comments and suggestions by Sybenga (1959), Bentzer et al. (1971) and Favarger (1978). For each taxon there is a presentation of the karyotype formula, maximum and minimum length of the chromosomes, total and average chromosome length and total haploid length of the chromosome set, along with their standard deviation. The interchromosomal asymmetry (CV<sub>CI</sub>), is estimated according to Paszko (2006) and the intrachromosomal asymmetry ( $M_{CA}$ ) according to Watanabe et al. (1999), Peruzzi and Eroğlu (2013) and Peruzzi and Altinordu (2014). Additionally the coefficient of variation of centromeric index (CV<sub>CI</sub>) measuring the centromere position heterogeneity is estimated following Paszko (2006) and Peruzzi and Altinordu (2014). A multivariate analysis (Principal Coordinate Analysis - PCoA) was made concerning six karyological parameters: 2n, x, THL,  $CV_{CI}$ ,  $CV_{CI}$  and  $M_{CA}$  (Peruzzi and Altinordu 2014). When marker chromosomes are observed (metacentric, submetacentric, SAT-chromosomes and secondary constrictions) r-index, R-length, Centromeric index and Arm difference ratio are also given. Finally, t-tests are given, regarding the indices of TCL, ACL,  $CV_{CL}$ ,  $M_{CA}$  in order to check statistically significant differences between the two species.

Taxon	Origin	2n	Voucher number, Herbarium		
F. montana	Mt. Vourinos (W Macedonia)	18	16765, UPA		
	Mt. Kato Olympos (Thessalia)	18	SF1089, ACA cult. no. 253, UPA		
	Mt. Boutsi (NW Macedonia)	27 and 18 (1 individual)	SF1092, ACA 19865, UPA		
F. epirotica	Katara Pass (Epirus)	24	21348, UPA 7919, UPA		
	Mt. Vasilitsa (N Pindos)	24	SF1076, ACA		
	Mt. Smolikas (N Pindos)	24	SF1097, ACA		
	Mt. Kratsovo (W Thessalia)	24	cult. no. 255, UPA		

Table 1. Origin, chromosome numbers (2n) and voucher number of *Fritillaria* material.

#### Results

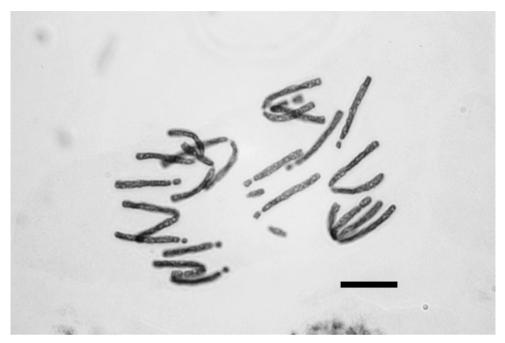
*Fritillaria montana* Hoppe — 2n = 2x = 18 + 0-3B (Figs 1, 2).

Populations karyologically studied:

Greece: Macedonia: Nomos Kozanis: mons Vourinos, in declivibus orientalibus cacuminis, alt. 1300-1350 m, in apertis ad viam et in silva Abietis et Pinetis, solo ophiolithico, 9 Jul 1981, T.R. Dudley, D. Phitos, D. Tzanoudakis, Gr. Iatrou & D. Christodoulakis 16765 (UPA); Thessalia: Nomos Larissis: Mt. Kato Olympos, Livadaki, north of Kallipefki, alt. ca. 1407 m, 39°57'N; 22°29'E, 30 May 2015, S. Samaropoulou, I. Patrikios & K. Tamvakas SF1089 (ACA); Mt. Kato Olympos, Livadaki, alt. 1400 m, May 2006, K. Tamvakas 253 (UPA).

*Fritillaria montana* is the only Greek species with a basic chromosome number of x = 9, having 2n = 18 chromosomes (Fig. 1). Its karyotype includes two metacentric (m) chromosome pairs that can be characterized as markers, the longer and the shorter ones (Table 3, chromosome pairs no. 1 and no. 5, numbered according to their chromosome length), because they bear characteristic secondary constrictions close to the end of the short arm (Fig. 2). Secondary constrictions are also observed to the rest of the metacetric chromosomes, however, they are not always visible. For this reason, the other three metacentric chromosome pairs cannot be characterized as markers.

The karyotype formula of the studied populations is given as 2n = 10m + 2st + 6t = 18 (Fig. 2). The chromosome size ranges between 24.41 µm and 11.26 µm and the total chromosome length is 316.34 µm. The karyotype is more symmetric (Table 2) concerning the variation in chromosome length ( $CV_{CL} = 25.2$ ) rather than the centromere position ( $M_{CA} = 41.42$ ), while the parameter  $CV_{CI}$  is even higher ( $CV_{CI} = 56.21$ ). Up to three B-chromosomes were found, all of them acrocentric (st) in the studied material.



**Figure 1.** Photomicrograph of mitotic metaphase plate of *Fritillaria montana* from Mt. Vourinos, 2n = 2x = 18. Bar = 10 µm.

**Table 2.** Studied species with karyomorphometric indices. Chromosome number (2n), total (TCL) and average (ACL) chromosome length, total haploid chromosome length (THL), maximum (max l + s) and minimum (min l + s) chromosome length, karyotype asymmetry indices ( $CV_{CL}$   $CV_{Cl}$  and  $M_{CA}$ ).

Species	F. me	F. epirotica			
Chromosome number	2n = 2x = 18	2n = 3x = 27	2n = 2x = 24		
Karyotype formula	10m + 2st + 6t	15m + 3st + 9t (10m + 4st + 4t, 1 individual)	2m + 2sm + 14st + 6t		
TCL (μm)	316.34	363.23	324.39		
(SD)	(30.22)	(53.47)	(51.12)		
THL (µm)	158.17	121.08	162.2		
(SD)	(15.11)	(17.82)	(25.56)		
ACL (µm)	17.57	13.45	13.52		
(SD)	(1.68)	(1.98)	(2.13)		
max l + s (μm)	24.41	22.86	18.44		
min l + s (μm)	11.26	8.00	10.00		
CV <sub>CL</sub>	25.26	31.07	16.85		
(SD)	(1.12)	(3.61)	(2.43)		
CV <sub>CI</sub> 56.21		54.79	51.66		
(SD) (0.99)		(3.57)	(2.99)		
M <sub>CA</sub>	41.42	40.41	63.33		
(SD)	(0.35)	(1.18)	(1.25)		



Figure 2. Karyogram of *Fritillaria montana* from Mt. Vourinos, 2n = 2x = 18. Bar = 10 µm.

**Table 3.** Karyomorphometric indices of marker chromosomes for each species, marker chromosome pairs (numbered according to their chromosome length), long arm's length (l), short arm's length (s), chromosome length (l + s) with minimum and maximum prices, r- index, Centromeric index, Arm difference ratio, R-length.

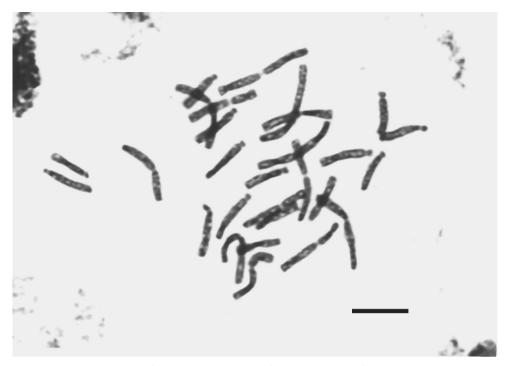
Species	Fritillaria	a montana	Fritillaria epirotica					
Chromosome number	2n =	= 18	2n = 24					
marker chromosomes	Pair no. 1	Pair no. 5	Pair no. 1	Pair no. 2	Pair no. 3	Pair no. 5		
l (μm)	12.84	10.71	9.68	10.51	12.19	10.87		
(SD)	(1.00)	(1.09)	(1.05)	(1.04)	(1.07)	(1.23)		
s (µm)	11.15	6.93	6.86	5,27	1.60	1,82		
(SD)	(0.79)	(0.67)	(0.82)	(0.63)	(0.44)	(0.36)		
l + s (µm)	23.99	17.65	16.53	15.77	13.79	12.42		
(SD)	(1.71)	(1.66)	(1.73)	(1.49)	(1.18)	(1.51)		
min l + s (µm)	20.88	15	13.53	12.94	11.47	8.40		
max l + s (µm)	26.47	20	19.12	18.24	15.59	15		
r-index l/s	1.15	1.54	1.40	2.03	8.06	6.24		
Centromeric index 1/1 + s	0.54	0.61	0.58	0.67	0.88	0.85		
Arm difference ratio l - s/l + s	0.70	0.21	0.17	0.34	0.77	0.71		
R-length l + s/Sn(l + s)	0.08	0.06	0.05	0.05	0.04	0.04		

*Fritillaria montana* Hoppe — 2n = 3x = 27 and 2n = 2x = 18 (1 individual) (Figs 3, 4).

Populations karyologically studied:

Greece: Macedonia: Nomos Florinas: Montes Triklarion, in declivibus boreooccidentalibus cacuminis Boutsi, in apertis saxosis calc., alt. 1450-1550 m, 19 May 1987, D. Phitos & G. Kamari 19865 (UPA); Mt. Boutsi, alpine meadow, calcareous substrate, alt. ca. 1549 m, 40°38'33"N; 21°09'25"E, 2 Jun 2015, S. Samaropoulou, I. Patrikios & A. Ioannou, sub Samaropoulou SF1092 (ACA).

The triploid population previously reported for the first time by Kamari (1991a), is now further examined. The karyotype formula is given as 2n = 15m + 3st + 9t = 27 (Figs 3, 4) and the chromosome length ranges from 22.86 µm to 8 µm, while the TCL equals to 363.23 µm (Table 2). The interchromosomal asymmetry of the triploid karyotype (CV<sub>CL</sub>



**Figure 3.** Photomicrograph of mitotic metaphase plate of *Fritillaria montana* from Mt. Boutsi, 2n = 3x = 27. Bar = 10  $\mu$ m.

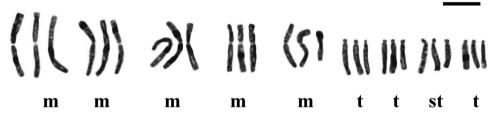


Figure 4. Karyogram of *Fritillaria montana* from Mt. Boutsi, 2n = 3x = 27. Bar = 10 µm.

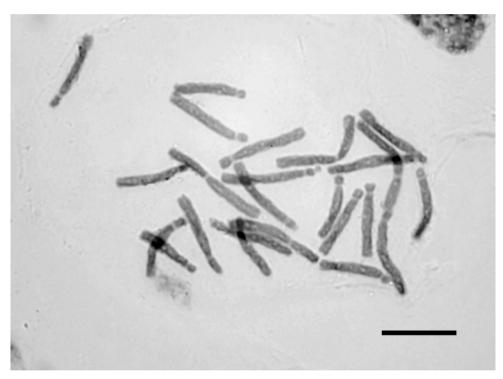
= 31.07) is higher than the diploid, but the intrachromosomal is lower ( $M_{CA}$  = 40.41). The heterogeneity of the centromere position is lower than the diploid ( $CV_{CI}$  = 54.79). Even though secondary constrictions were observed again, their number and position varies in several plates, making the distinction of marker chromosomes very difficult.

It is noteworthy that a diploid individual was found for the first time at the studied triploid population. The karyotype of this individual comprises 2n = 10m + 4st + 4t = 18 chromosomes, with an additional pair of acrocentric (st) chromosomes compared with the other diploid populations studied and without B-chromosomes in contrast with the population of Mt. Vourinos. The secondary constrictions were also unclear as in the triploid individuals.

#### *Fritillaria epirotica* Turrill ex Rix — 2n = 2x = 24 (Figs 5, 6).

Populations karyologically studied:

Greece: Epirus: Nomos Ioanninon: Katara Pass, prope ekchionistikos stathmos, alt. 1750 m, in apertis (*Pinus* Linnaeus, 1753; *Buxus* Linnaeus, 1753 etc), solo serpentinico, 4 May 1990, D. Phitos & G. Kamari 21348 (UPA); Eparchia Metsovou, Katara Pass, close to the second snowplow station, c. 13.5 km of Metsovon along the road to Trikala, slopes with *Pinus nigra* Arnold, 1785 and *Buxus sempervirens* Linnaeus,1753; ophiolithic substrate, alt. c. 1640 m, 39°47'N; 21°13'E, 24 Jun 1998, Th. Constantinidis 7919 (UPA); Macedonia: Nomos Grevenon: Mt. Vasilitsa, alt.



**Figure 5.** Photomicrograph of mitotic metaphase plate of *Fritillaria epirotica* from Mt. Smolikas, 2n = 2x = 24. Bar = 10  $\mu$ m.



**Figure 6.** Karyogram of *Fritillaria epirotica* from Mt. Smolikas, 2n = 2x = 24. Bar = 10 µm.

1764 m, 17 May 2015, G. Kofinas s.n. (cult. no. SF76, ACA); Mt. Smolikas, alt. 2200 m, Aug 2015, G. Kofinas s.n. (cult. no. SF97, ACA). **Thessalia: Nomos Trikalon:** Ep. Kalampakas, Mt. Chasia (Kratsovo), stony slopes close to a forest road, c. 3.0–3.5 km from Kakoplevri village, serpentine, alt. c. 1100–1180 m, 39°48'N; 21°24'E, 15 Jun 2000, D. Phitos, G. Kamari & Th. Constantinidis s.n. (cult. no. 235, UPA); Ep. Kalampakas, Mt. Chasia (Mt. Kratsovon), c. 3.1 km WNW of Kakoplevri village on the foothills of the mountain, hills with low *Buxus sempervirens* and *Juniperus oxycendrus* Linnaeus,1753; serpentine substrate, alt. 1120–1160 m, 39°49'N; 21°24'E, 24 Jul 2006, Th. Constantinidis s.n. (cult. no. 235, UPA).

Unlike *Fritillaria montana*, *F. epirotica* has the same basic somatic number as the rest of the Greek *Fritillaria* taxa, x = 12. The karyotype consists of 2n = 2m + 2sm + 14st + 6t = 24 chromosomes (Figs 5, 6), which range in size between 18.44 and 10 µm, while the TCL is 324.39 µm (Table 2). The index for interchromosomal asymmetry is small ( $CV_{CL} = 16.85$ ) contradicting the big intrachromosomal one ( $M_{CA} = 63.33$ ), while the centromere position heterogeneity is 51.66. Satellites on the short arms of one telocentric (t) and one acrocentric (st) pair of chromosomes (Table 3, chromosome pairs no. 3 and no. 5) are observed. However, in most metaphase plates, three of them are usually visible.

According to paired t-tests made (Table 4), the two species display an interesting similarity regarding their total chromosome length, but as far as the interchromosomal and intrachromosomal asymmetries are concerned (Table 5), the species seem to be

**Table 4.** Paired t-tests between the three species regarding the TCL and ACL along with degrees of freedom (df) and Significance (Sig) for every parameter. Bold characters are used for P values (Sig 2-tailed) under 0.01, which reveal significant statistical difference.

Species in comparison		TCL			ACL			
		t	df	Sig (2-tailed)	t	df	Sig (2-tailed)	
F. epirotica	F. montana	0.379	18	0.709	-4.347	18	0.000	
2n = 2x = 24	2n = 2x = 18	0.577	10	0.707	-1.51/	10	0.000	
F. epirotica	F. montana	1 427	-1.427 10	16	0.173	0.057	16	0.955
2n = 2x = 24	2n = 3x = 27	-1.42/	-1.42/ 10	0.1/3	0.057	10	0.775	
F. montana	F. montana	-1.947	10	0.080	3.898	10	0.003	
2n = 2x = 18	2n = 3x = 27	-1.94/	10	0.080	3.090	10	0.005	

**Table 5.** Paired t-tests between the three species regarding the  $CV_{CL}$  and  $M_{CA}$ , along with degrees of freedom (df) and Significance (Sig) for every parameter. Bold characters are used for P (Sig 2-tailed) under 0.01, which reveal significant statistical difference.

Species in comparison		CV <sub>CL</sub>			M <sub>CA</sub>		
		t	df	Sig (2-tailed)	t	df	Sig (2-tailed)
F. epirotica	F. montana	-8.598	18	0.000	44.847	18	0.000
2n = 2x = 24	2n = 2x = 18	-0.990	10	0.000	44.04/	10	0.000
F. epirotica	F. montana	-9.754	16	0.000	34.473	16	0.000
2n = 2x = 24	2n = 3x = 27						
F. montana	F. montana	-4.066	6 10	0.002	1.005	10	0.074
2n = 2x = 18	2n = 3x = 27			0.002	1.995	10	0.074

clearly distinct. The only insignificant difference was revealed between the two cytotypes of *Fritillaria montana*, 2n = 18 and 2n = 27, as expected, since they both bear a lot of metacentric chromosomes (by Robersonian fusions).

#### Discussion

In the present study a detailed karyomorphological analysis of *Fritillaria montana* and *Fritillaria epirotica*, in material from Greece, was implemented focusing specifically to the study of the inter- and intrachromosomal asymmetry, as well as the detailed analysis of the marker chromosomes.

The study of marker chromosomes (Table 3) is always important since it can provide further information concerning genome organization and the differentiation of the karyotype between related species. Moreover, especially in the case of the genus *Fritillaria*, marker chromosomes are helpful for the distinction of the chromosome homologues, which is very difficult since the karyotype usually consists of mostly acrocentric and subtelocentric chromosomes with similar size.

Marker chromosomes were observed in both *F. epirotica* with 2n = 2x = 24 and *F. montana* with 2n = 2x = 18 chromosomes. However, when it comes to triploid karyotypes of the same species, the secondary constrictions are not stable in number and position.

*Fritillaria epirotica* (2n = 24) has four marker chromosome pairs (Fig. 6). The first two chromosome pairs, which are the longest ones of the complement, have a different morphology than all the other chromosomes of the karyotype, which are acrocentric (st) and subtelocentric (t). The longest chromosome pair is metacentric (m) (no. 1), while the second one is the second in range of length and a submetacentric (sm) one (no. 2). The third marker chromosome pair (no. 3) is telocentric and bears small spherical satellite on the short arm of the homologues. Finally, the last marker chromosome pair is the fifth in length, comprising of two acrocentric satellited (st-SAT) chromosomes. The results are in agreement with previous studies by Kamari (1991a). Zaharof (1989) reported the heteromorphism of satellites' length in one out of two SAT-chromosome pairs.

*Fritillaria montana* (2n = 18) has two marker chromosome pairs with secondary constrictions. The karyotype formula given here (2n = 10m + 2st + 6t = 18) differs from the previously reported karyotype of 2n = 10m + 8t = 18 chromosomes given by Zaharof (1989). This is the only one species in Greece with 18 chromosomes and this chromosomal reduction has already been claimed as the result of successive chromosomal reconstructions and Robersonian-fusion of six acrocentric chromosomes into three metacentric ones (Darlington 1936, La Cour 1978a, 1978b, 1978c, Kamari 1991a). Zaharof (1989) explained the secondary constrictions, which are also observed in the present study, with the above hypothesis. Recently, Peruzzi et al. (2016) studied an Italian population with 2n = 2x = 18 chromosomes, further confirming the chromosome number of *F. montana*, while the presence of up to three B-chromosomes is already referred by Kamari (1991a, 1991b).

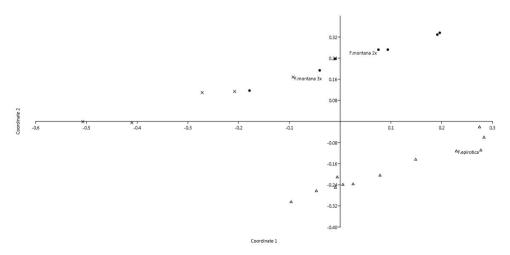
The triploid chromosome number of *F. montana* (2n = 3x = 27) is known in Greece from only one population, but it has also been reported from Italy by Cesca (1986, under the name *F. tenella* Marschall von Bieberstein, 1808), for a Calabrian population (S Italy).

Paired t-tests have revealed similarities among the three karyotypes. Especially the similarity between TCL of the diploid *F. epirotica* 2n = 24 and *F. montana* 2n = 18 reinforces the hypothesis, apart from the secondary constrictions, that the second species has derived after successive chromosomal reconstructions and Robersonian-fusions. Less similar indices of TCL between *F. montana* 2n = 2x = 18 and *F. montana* 2n = 3x = 27 can also be explained since it is known that polyploidy usually comes with gene loss and genome amount reduction (Kamari 1992, Leitch and Bennett 2004, Adams and Wendel 2005, Buggs et al. 2009). Another proof for gene loss, is the fact that the triploid cytotype of *F. montana* has the lower price of THL.

The results concerning the heterogeneity of centromere position  $CV_{CI}$  and the intrachromosomal asymmetry  $M_{CA}$  are nothing but expected. Following the explanation of this index by Zuo and Yuan (2011), the higher price of  $CV_{CI}$  found here belongs to *Fritillaria montana*, because the karyotype comprises of mostly metacentric chromosomes. On the contrary, the higher price of  $M_{CA}$  belongs to *F. epirotica*, as it has a typical asymmetrical karyotype according to Stebbins (1971).

In total, the multivariate analysis PCoA confirms all above findings. More precisely, it presents all the accessions belonging to the same species close to each other. The two cy-totypes of *Fritillaria montana* overlap, while the two species are clearly separated (Fig. 7).

Generally, karyological characteristics, as chromosome number, ploidy level, centromere position, and the number and location of satellites and secondary constrictions, can be used in elucidating taxonomical relationships of several plant taxa (Bareka et al. 2008, 2012 see for references). Although, karyomorphometrics is able



**Figure 7.** PCoA analysis based on six quantitative karyologial parameters. Triangle depicts *F. epirotica*, 2n = 2x = 24; dots *F. montana*, 2n = 2x = 18 and x *F. montana*, 2n = 3x = 27.

to provide more information about the studied taxa, the conclusions can be used only as additional evidences to the primary hypothesis. However, molecular chromosomal markers and fluorescence in situ hybridization (FISH) could provide additional information concerning genome organization in the genus and differentiation among its species and are recommended as a safer way to reveal whether our assumption for the origin of the reduced chromosome number 2n = 18 is correct. Moreover, this method is desirable to be carried out because it will unveil the type of polyploidy for 2n = 3x =27, as an autopolyploidy or allopolyploidy (Bareka et al. 2012).

### References

- Adams KL, Wendel JF (2005) Polyploidy and genome evolution in plants. Current Opinion in Plant Biology 8: 135–141. doi: 10.1016/j.pbi.2005.01.001
- Advay M, Tekşen M, Maroofi H (2015) *Fritillaria avromanica* sp. nov. (Liliaeceae) from Iran and notes on *F. melananthera* in Turkey. Nordic Journal of Botany 33: 526–531. doi: 10.1111/njb.00780
- Ambrozova K, Mandakova T, Bures P, Neumann P, Leitch IJ, Koblızkova A, Macas J, Lysak MA (2011) Diverse retrotransposon families and an AT-rich satellite DNA revealed in giant genomes of *Fritillaria* lilies. Annals of Botany 107: 255–268. doi: 10.1093/aob/mcq235
- Bareka P, Phitos D, Kamari G (2008) A karyosystematic study of the genus *Bellevalia* Lapeyr. (Hyacinthaceae) in Greece. Botanical Journal of the Linnean Society 157: 723–739. doi: 10.1111/j.1095-8339.2008.00817.x
- Bareka P, Siljak-Yakovlev S, Kamari G (2012) Molecular cytogenetics of *Bellevalia* (Hyacinthaceae) species occurring in Greece. Plant Systematics Evolution 298: 421–430. doi: 10.1007/s00606-011-0555-7
- Bartolucci F, Caparelli KF, Peruzzi L (2009) A biometric study of *Fritillaria montana* Hoppe s.l. (Liliaceae) shows a single polymorphic species, with no infraspecific taxa. Plant Biosystems 143(3): 516–527. doi: 10.1080/11263500902722956
- Bennett MD, Smith JB (1976) Nuclear DNA amounts in angiosperms. Philosophical Transactions of the Royal Society B: Biological Sciences 274: 227–274. doi: 10.1098/ rstb.1976.0044
- Bentzer B, Bothmer R von, Engstrand L, Gustafsson M, Snogerup S (1971) Some sources of error in the determination of arm ratios of chromosomes. Botaniska Notiser 124: 65–74.
- Buggs RJA, Doust AN, Tate JA, Koh J, Soltis K, Feltus FA, Paterson AH, Soltisand PS, Soltis DE (2009) Gene loss and silencing in *Tragopogon miscellus* (Asteraceae): comparison of natural and synthetic allotetraploids. Heredity 103: 73–81. doi: 10.1038/hdy.2009.24
- Cesca G (1986) Note fitogeografiche e citotassonomiche su: *Adoxa moschatellina* L., *Tulipa sylvestris* L., *Fritillaria tenella* Bieb. Biogeographia (N.S) 10 (1984): 109–139.
- Chorney II, Kagalo AA, Lyubinska LG (2009) *Fritillaria montana* Hoppe. In: Diduh Ya.P. (Ed.) Red Book of Ukraine. Plant world, Globalconsulting, Kyiv, 139.
- Conti F, Manzi A, Pedrotti F (1997) Liste Rosse regionali delle piante d'Italia. Camerino: WWF and Societa Botanica Italiana. Camerino, Italy, 139 pp.

- Darlington CD (1935) The internal mechanics of the chromosomes I. The nuclear cycle in *Fritillaria*. Proceedings of the Royal Society of London-Series B 118: 33–59. doi: 10.1098/ rspb.1935.0047
- Darlington CD (1936) The external mechanics of chromosomes. Proceedings of the Royal Society of London Biological Sciences 121: 264–319. doi: 10.1098/rspb.1936.0064
- Darlington CD (1937) Recent Advances in Cytology. Churchill, London, UK, 671 pp.
- Darlington CD, La Cour LF (1969) The Handling of Chromosomes. London, UK, 272 pp.
- Favarger C (1978) Philosophie des comptages de chromosome. Taxon 27: 441–448. doi: 10.2307/1219893
- Fay MF, Chase MW (2000) Modern concepts of Liliaceae with a focus on the relationships of *Fritillaria*. Curtis's Botanical Magazine 17: 146–149. doi: 10.1111/1467-8748.00258
- Fedorov A (1969) Chromosome numbers of flowering plants. Leningrad, 926 pp. [In Russian]
- Frankel OH (1940) The causal sequence of meiosis. I. Chiasma formation and the order of pairing in *Fritillaria*. Journal of Genetics 41: 9–34. doi: 10.1007/BF02982973
- IUCN (2014) IUCN Red List Categories and Criteria: version 2014.2. IUCN Species Survival. Commission, Gland & Cambridge.
- Jafari H, Babaei A, Karimzadeh G (2014) Cytogenetic study on some *Fritillaria* species o Iran. Plant Systematics and Evolution 300: 1373–1383. doi: 10.1007/s00606-013-0968-6
- Kamari G (1976) Cytotaxonomic study of the *Crepis neglecta* L. complex in Greece. Ph.D. Thesis, University of Patras, Patras, Greece, 193 pp. [In Greek]
- Kamari G (1984a) Caryosystematic studies of *Fritillaria* L. (Liliaceae) in Greece. 1. Webbia 38: 723–731. doi: 10.1080/00837792.1984.10670343
- Kamari G (1984b) *Fritillaria sporadum* (Liliaceae), a new species from N. Sporades (Greece). Willdenowia 14: 331–333.
- Kamari G (1991a) The genus *Fritillaria* L. in Greece: taxonomy and karyology. Botanika Chronika 10: 253–270.
- Kamari G (1991b) Fritillaria L. In: Strid A, Tan K (Eds) Mountain flora of Greece, 2, Edinburgh University Press, Edinburgh, UK, 672–683.
- Kamari G (1992) Karyosystematic studies on three *Crepis* species (Asteraceae) endemic to Greece. Plant Systematics and Evolution 182: 1–19. doi: 10.1007/BF00941411
- Kamari G (1995) Fritillaria epirotica Turill ex Rix. In: Phitos D, Strid A, Snogerup S, Greuter W (Eds) The Red Data Book of Rare and Threatened Plants of Greece, WWF Hellas, Athens, Greece, 272–273.
- Kamari G (1996) *Fritillaria* species (Liliaceae) with yellow or yellowish-green flowers in Greece. Bocconea 5: 221–238.
- Kamari G, Phitos D (2000) *Fritillaria theophrasti*, a new species from Lesvos, East Aegean islands, Greece. Biologia Gallo-hellenica, Supplementum 26: 69–76.
- Kamari G, Phitos D (2006) Karyosystematic study of *Fritillaria messanensis* s.l. (Liliaceae). Willdenowia 36: 217–233. doi: 10.3372/wi.36.36118
- Kamari G, Phitos D (2009) Fritillaria epirotica Turill ex Rix. In: Phitos D, Constantinidis Th, Kamari G (Eds) The Red Data Book of Rare and Threatened Plants of Greece, Vol. II (E-Z), Hellenic Botanical Society, Patras, Greece, 37–38.

- Koul KA, Wafai BA (1980) Chromosome polymorphism and nucleolar organization in some species of *Fritillaria*. Cytologia 45: 675–682. doi: 10.1508/cytologia.45.675
- La Cour LF (1978a) The constitutive heterochromatin in the chromosomes of *Fritillaria* sp., as revealed by Giemsa banding. Philosophical Transactions of the Royal Society B: Biological Sciences 285(1004): 61–71. doi: 10.1098/rstb.1978.0094
- La Cour LF (1978b) Notes and comments. Differential Giemsa staining of B-chromosomes of *Fritillaria tenella*. Heredity 41: 101–103. doi: 10.1038/hdy.1978.68
- La Cour LF (1978c) Two types of constitutive heterochromatin in the chromosomes of some *Fritillaria* species. Chromosoma 67: 67–75. doi: 10.1007/BF00285649
- Leitch IJ, Bennett MD (2004) Genome downsizing in polyploidy plants. Biological Journal of the Linnean Society 82: 651–663. doi: 10.1111/j.1095-8312.2004.00349.x
- Levan A, Fredga K, Sandberg A (1965) Nomenclature for centromeric position on chromosomes. Hereditas 52: 201–220. doi: 10.1111/j.1601-5223.1964.tb01953.x
- Li R, Shang Z (1989) The chromosome observation on five species of rare plants of China. Journal of Wuhan Botanical Research 7: 217–220.
- Lozina-Lozinskaja AS (1935) Fritillaria L. In: Komarov VL (Ed.) Flora URSS, 4, USSR Academy of Sciences, Leningrad, 302–320. [In Russian].
- Mancuso E, Bedini G, Peruzzi L (2012) Morphology, germination and storage behaviour in seeds of *Fritillaria montana* (Liliaceae) a rare perennial geophyte in Italy. Turkish Journal of Botany 36: 161–166.
- Marchant CJ, Macfarlane RM (1980) Chromosome polymorphism in triploid populations of *Fritillaria lanceolata* Pursh (Liliaceae) in California. Botanical Journal of the Linnean Society 81: 135–154. doi: 10.1111/j.1095-8339.1980.tb00945.x
- Mehra PN, Sachdeva SK (1976) Cytological observations on some W. Himalayan monocots. II. Smilacaceae, Liliaceae and Trilliaceae. Cytologia 41: 5–22. doi: 10.1508/cytologia.41.5
- Metin OK, Turktas M, Aslay M, Kaya E (2013) Evaluation of the genetic relationship between *Fritillaria* species from Turkey's flora using fluorescent-based AFLP. Turkish Journal of Botany 37: 273–279.
- Moore DM (1982) Flora europaea. Checklist and chromosome index. Cambridge, UK, 423 pp.
- Noda S (1964) Cytology in the genus *Fritillaria*. I. Variations in karyotypes and B-chromosomes in *F. amabilis*. Bulletin of the Osaka Gakuin University 2: 125–132.
- Östergren G, Heneen WK (1962) A squash technique for chromosome morphological studies. Hereditas 48: 332–341. doi: 10.1111/j.1601-5223.1962.tb01817.x
- Özhatay N (2000) *Fritillaria* L. In: Güner A, Özhatay N, Ekim T, Başer KHC (Eds) Flora of Turkey and the East Aegean Islands (Supp. 2), 11. Edinburgh University Press. Edinburgh, UK, 243–246.
- Özhatay N, Wallis R, Wallis RB, Kocyiğit M (2015) A new *Fritillaria* species from Mediterranean region of Turkey; *Fritillaria asumaniae*. Flora Mediterranea 25 (Special Issue): 199–208.
- Paszko A (2006) A critical review and a new proposal of karyotype asymmetry indices. Plant Systematics and Evolution 258: 39–48. doi: 10.1007/s00606-005-0389-2
- Peruzzi L, Mancuso E, Ansaldi M, De Angeli E, Trombetti G (2008) Distribuzione, caratterizzazione e consistenza delle popolazioni toscane di *Fritillaria montana* Hoppe (Liliaceae).
  Webbia 63: 309–315. doi: 10.1080/00837792.2008.10670846

- Peruzzi L, Bartolucci F (2009) Typification of the names within *Fritillaria montana* complex (Liliaceae) from central Mediterranean area. Candollea 64(1): 133–142.
- Peruzzi L, Leitch IJ, Caparelli KF (2009) Chromosome diversity and evolution in Liliaceae. Annals of Botany (London) 103: 459–475. doi: 10.1093/aob/mcn230
- Peruzzi L, Mancuso E, Gargano D (2012) Males are cheaper, or the extreme consequence of size/ age dependent sex allocation: sexist gender diphasy in *Fritillaria montana* (Liliaceae). Botanical Journal of the Linnean Society 168(3): 323–333. doi: 10.1111/j.1095-8339.2011.01204.x
- Peruzzi L, Eroğlu HE (2013) Karyotype asymmetry: again, how to measure and what to measure? Comparative Cytogenetics 7(1): 1–9. doi: 10.3897/compcytogen.v7i1.4431
- Peruzzi L, Altinordu F (2014) A proposal for a multivariate quantitative approach to infer karyological relationships among taxa. Comparative Cytogenetics 8(4): 337–349. doi: 10.3897/CompCytogen.v8i4.8564
- Peruzzi L, Astuti G, Bartolucci F, Conti F, Rizzotto M, Roma-Marzio F (2016) Chromosome numbers for the Italian flora. Italian Botanist I: 39–53. doi: 10.3897/italianbotanist.1.8818
- Petrova A, Vladimirov V (2009) Red List of Bulgarian vascular plants. Phytologia Balcanica 15(1): 63–94.
- Phitos D, Strid A, Snogerup S, Greuter W (1995) The Red Data Book of Rare and Threatened Plants of Greece. WWF Hellas. Athens, Greece, 527 pp.
- Phitos D, Constantinidis Th, Kamari G (2009) The Red Data Book of Rare and Threatened Plants of Greece, Vol. II (E-Z). Hellenic Botanical Society. Patras, Greece, 413 pp.
- Rix EM (1977) Fritillaria L. (Liliaceae) in Iran. Iranian Journal of Botany 1(2): 75-95.
- Rix EM (1984) *Fritillaria* L. In: Davis PH (Ed.) Flora of Turkey and the East Aegean Islands,
  8. Edinburgh University Press, Edinburgh, UK, 284–302.
- Rix EM (2001) Fritillaria. A Revised Classification. The Fritillaria Group of the Alpine Garden Society, United Kingdom, 14 pp.
- Rønsted N, Law S, Thornton H, Fay MF, Chase MW (2005) Molecular phylogenetic evidence for the monophyly of *Fritillaria* and *Lilium* (Liliaceae; Liliales) and the infrageneric classification of *Fritillaria*. Molecular Phylogeny and Evolution 35: 509–527. doi: 10.1016/j. ympev.2004.12.023
- Runemark H (1970) The role of small populations for the differentiation in plants. Taxon 19: 196–201. doi: 10.2307/1217954
- Sharma S, Raina SN (2005) Organization and evolution of highly repeated satellite DNA sequences in plant chromosomes. Cytogenetic and Genome Research 109: 15–26. doi: 10.1159/000082377
- Šilić Č (1996) The List of Plant Species (Pteridophyta and Spermatophyta) for the Red Book of Bosnia and Herzegovina. Glasnik Zemaljskog Muzeja Bosne i Hercegovine, Sarajevo 31: 323–367.
- Stebbins GL (1971) Chromosomal evolution in higher plants. Edward Arnold Ltd., London, UK, 220 pp.
- Sybenga J (1959) Some sources of error in the determination of chromosome length. Chromosoma 10: 355–364. doi: 10.1007/BF00396578
- Tekşen M, Aytaç Z (2011) The revision of the genus *Fritillaria* L. (Liliaceae) in the Mediterranean region (Turkey). Turkish Journal of Botany 35: 447–478.

- Tekşen M (2012) *Fritillaria* L. In: Güner A, Aslan S, Ekim T, Vural M, Babaç MT (Eds) Türkiye Bitkileri Listesi (Damarlı Bitkiler). Nezahat Gökyiğit Botanik Bahcesi ve Flora Araştırmaları Derneği Yayını, Turkey, 604–607.
- Tomović G, Vukojić S, Niketić M, Zlatiković B, Stevanović V (2007) *Fritillaria* (Liliaceae) in Serbia: distribution, habitats and some taxonomic notes. Phytologia Balcananica 13(3): 359–370.
- Türktaş M, Aslay M, Kaya E, Ertuğrul F (2012) Molecular characterization of phylogenetic relationships in *Fritillaria* species inferred from chloroplast trnL trnF sequences. Turkish Journal of Botany 36: 552–560.
- Velchev V (Ed.) (1984) Red Data Book of the PR Bulgaria, 1. Plants. Publishing House Bulgarian Academy of Sciences, Sofia, 448 pp.
- Watanabe K, Yahara T, Denda T, Kosuge K (1999) Chromosomal evolution in the genus Brachyscome (Asteraceae, Astereae): Statistical tests regarding correlation between changes in karyotype and habit using phylogenetic information. Journal of Plant Research 112: 145–161. doi: 10.1007/PL00013869
- Zahariadi C (1966) Liliaceae AL Jussieu. In: Săvulescu T (Ed.) Flora Reipublicae Socialisticae România 9, Editio Academiae Reipublicae Socialisticae Romania, Bucharest, 106–404.
- Zaharof E (1987) Biometrical and cytological studies of the genus *Fritillaria* L. in Greece. Ph.D. Thesis, University of Thessaloniki, Greece, 236 pp. [In Greek]
- Zaharof E (1989) Karyological studies of twelve *Fritillaria* species from Greece. Caryologia 43: 91–102. doi: 10.1080/00087114.1989.10796956
- Zuo J, Yuan G (2011) The difference between the heterogeneity of the centromeric index and intrachromosomal asymmetry. Plant Systematics and Evolution 297: 141–145. doi: 10.1007/s00606-011-0528-x