RESEARCH ARTICLE



# Karyotypes of six spider species belonging to the families Gnaphosidae, Salticidae, Thomisidae, and Zodariidae (Araneae) from Turkey

Zübeyde Kumbıçak<sup>1</sup>, Emel Ekiz<sup>2</sup>, Serdar Çiçekli<sup>2</sup>

l Department of Molecular Biology and Genetics, Faculty of Science and Art, Nevşehir Hacı Bektaş Veli University, 50300, Nevşehir, Turkey **2** Department of Biology, Faculty of Science and Art, Nevşehir Hacı Bektaş Veli University, 50300, Nevşehir, Turkey

Corresponding author: Zübeyde Kumbıçak (akanzubeyde@gmail.com)

Academic editor: Jiří Král | Received 6 August 2013 | Accepted 12 March 2014 | Published 16 May 2014

**Citation:** Kumbıçak Z, Ekiz E, Çiçekli S (2014) Karyotypes of six spider species belonging to the families Gnaphosidae, Salticidae, Thomisidae, and Zodariidae (Araneae) from Turkey. Comparative Cytogenetics 8(2): 93–101. doi: 10.3897/ CompCytogen.v8i2.6065

#### Abstract

In this study, the karyotypes of six spider species from Turkey belonging to the families Gnaphosidae, Salticidae, Thomisidae, and Zodariidae were analyzed. Male chromosomal features including diploid chromosome numbers and sex chromosome systems were determined as 2n=22,  $X_1X_20$  in *Drassyllus sur* Tuneva & Esyunin, 2003, *Nomisia exornata* (C. L. Koch, 1839), and *Nomisia orientalis* Dalmas, 1921; 2n=28,  $X_1X_20$  in *Sitticus caricis* (Westring, 1861); 2n=23, X0 in *Xysticus gallicus* Simon, 1875 and 2n=42,  $X_1X_20$  in *Pax islamita* (Simon, 1873), respectively. The chromosome morphology of all species was acrocentric. Data obtained contribute to knowledge of the karyotype evolution of araneomorphs.

#### Keywords

Araneae, diploid number, sex chromosome system

# Introduction

Spiders are one of the most important animal groups, and contain approximately 44 500 species all around the world (Platnick 2014) and consists of three primary clades, namely Mesothelae, Mygalomorphae and Araneaomorphae, the last one being phylogenetically most derived and the largest group (Coddington and Levi 1991). Infraorder

Araneaomorphae contains more than 41 000 species (Platnick 2014). Despite this diversity, only 771 species of spiders have been karyotyped so far (Araújo et al. 2014).

Entelegyne spiders form a very diversified clade of araneomorphs. Their karyotypes are characterized by a predominance of acrocentric chromosomes,  $X_1X_20$  sex chromosome system (Araújo et al. 2005), relatively low diploid chromosome numbers (ranges from 10 to 49, Kořínková and Král 2013), and chiasmatic meiosis (Kumbıçak 2010). Acrocentric karyotypes of entelegynes with lower chromosome numbers could be derived from ancestral acrocentric karyotypes by tandem fusions (Suzuki 1954) or by cycles of centric fusions and subsequent pericentric inversions (Kořínková and Král 2013). The latter hypothesis is supported by the fact that centric fusions are the most frequent source of chromosome polymorphism found in populations of entelegyne spiders (Kořínková and Král 2013).

In spiders, the  $X_1X_20$  system could be the ancestral sex chromosome determination as inferred from its presence in the most primitive recent spiders, namely the suborder Mesothelae and basal families of the infraorder Mygalomorphae (Suzuki 1954).

Salticidae, Thomisidae, Gnaphosidae, and Zodariidae are some of the largest families in the order Araneae (Platnick 2014). Despite the high diversity of these spider groups, cytogenetic data have been collected only in 223 species belonging to these clades (Araújo et al. 2014).

This study presents karyotypes of six species belonging to the genera *Drassyllus* Chamberlin, 1922 and *Nomisia* Dalmas, 1921 (Gnaphosidae), *Sitticus* Simon, 1901 (Salticidae), *Xysticus* C. L. Koch, 1835 (Thomisidae), and *Pax* Levy, 1990 (Zodariidae). Our study brings new data and fills some gaps in cytogenetics of these families.

## Material and methods

**Material**: Spiders were collected in Mediterranean, Southeast and Central Anatolia (Turkey) during the year 2012. Collection data of particular species (localities including their coordinates, dates of collection, number of individuals studied) are listed in Table 1. Voucher specimens were deposited in the collection of Department of Molecular Biology and Genetics, Art and Science Faculty, Nevşehir Hacı Bektaş Veli University (Nevşehir, Turkey). The identification of spiders was made by O. Seyyar (Department of Biology, Art and Science Faculty, Niğde University, Niğde, Turkey).

**Chromosome preparations and observation:** Slides for chromosome observations were made by the spreading technique of Traut (1976), with some modifications. This method consisted of three basic steps. First, the gonads were hypotonized in 0.075 M KCl for 12-15 min in room temperature (RT). Second, gonads were fixed in two batches of freshly prepared Carnoy fixative (ethanol: chloroform: glacial acetic acid; 6:3:1), first batch for 10 min and second one for 20 min (RT). Finally, a cell suspension was prepared from a piece of tissue in a drop of 60% acetic acid on a slide using a pair of tungsten needles. The slide was placed on a histological plate at 42 °C and the drop was evaporated by mov-

Family	Species	Locality	Coordinates	Date of Collection	Number of Individuals Studied
Gnaphosidae	<i>Drassyllus sur</i> Tuneva & Esyunin, 2003	Gaziantep, Sakçagözü	37°10'18"N, 36°55'39"E	04.04.2012	7්
	<i>Nomisia exornata</i> (C. L. Koch, 1839)	Antalya, Aksu	36°55'30"N, 30°48'29"E	24.03.2012	11ð
		Antalya, Gazipaşa	36°16'23"N, 32°17'33"E	24.03.2012	2්
	Nomisia orientalis	Osmaniye, Düziçi	37°15'02"N, 36°26'36"E	21.05.2012	58
	Dalmas, 1921	Adıyaman, Kahta	37°48'46"N, 38°38'20"E	11.03.2012	48
		Gaziantep, Islahiye	37°01'21"N, 36°37'24"E	06.04.2012	9♂
Salticidae	Sitticus caricis	Nevşehir, Göreme	38°38'44"N, 34°50'06"E	10.05.2012	8්
	(Westring, 1861)	Nevşehir, Zelve	38°40'16"N, 34°51'43"E	27.06.2012	3්
Thomisidae	V II.	Adana, Çamalan	37°19'12"N, 34°36'28"E	12.04.2012	6්
	<i>Xysticus gallicus</i>	Mersin, Bozyazı	36°06'04"N, 32°58'38"E	15.04.2012	2්
	Simon, 1875	Mersin, Aydıncık	36°08'36"N, 33°22'59"E	15.04.2012	3්
Zodariidae	Pax islamita (Simon, 1873)	Osmaniye, Toprakkale	37°04'24"N, 36°08'42"E	09.06.2012	5්

**Table 1.** Material used for chromosome analysis.

ing it with a tungsten needle. Slides were stained with 5% Giemsa in Sørensen phosphate buffer (pH=6.8) for 27 min (RT). Chromosome spreads were investigated an Olympus BX53 microscope and photographed using a DP26 digital camera (Olympus) using CELLSENS software (Olympus). Relative chromosome lengths (RCL) including standard deviations were calculated as a percentage of the total chromosome length of the diploid set including sex chromosomes (%TCL) from 10 mitotic metaphase plates for each species by CELLSENS software. Classification of chromosome morphology was based on the arm ratio (Levan et al. 1964).

#### Results

#### Gnaphosidae

The chromosomes of *Drassyllus sur* Tuneva & Esyunin, 2003 (2n = 22) were acrocentric. The sex chromosome system was formed by chromosomes  $X_1$  and  $X_2$  which were medium-sized elements (Fig. 1A). The autosome pairs decreased gradually in size. Length of autosome pairs decreased from 9.74±0.29% to 6.89±0.12% of TCL. Relative length of  $X_1$  and  $X_2$  was 8.45±0.06% and 7.57±0.17% of the diploid set, respectively.

There were 10 autosomal bivalents and two sex chromosomes at diplotene (Fig. 2A). Sex chromosomes were positively heteropycnotic from leptotene to metaphase II (Fig. 2B).

<b>fc</b> A 1	<b>11</b> 2	<b>K</b> 3	<b>)(</b> 4	<b>()</b> 5	<b>(1</b> 6	<b>(</b> c 7	<b>N</b> 8	<b>11</b> 9	<b>11</b>				X1 X2
<b>cl</b> B 1	<b>\$c</b> 2	<b>X</b> 3	<b>)</b> ( 4	<b>11</b> 5	<b>11</b> 6	<b>s</b> t 7	<b>11</b> 8	9	<b>II</b>				X1 X2
С 1	<b>11</b> 2	<b>H</b> 3	<b>((</b> 4	5	<b>()</b> 6	<b>))</b> 7	<b>1</b> ( 8	<b>N</b> 9	<b>10</b>				$X_1$ $X_2$
<b>C</b> ( D 1		<b>r</b> 3	<b>۲</b> 4	<b>C</b> 5	<b>)</b> C 6	(2 7	<b>\$</b> 5 8	<b>75</b>	<b>(</b> ) 10	<b>c(</b> 11	<b>L(</b> 12	<b>1</b> 3	$\begin{array}{c} \mathbf{\zeta} & \mathbf{\zeta} \\ \mathbf{X}_1 & \mathbf{X}_2 \end{array}$
<b>к</b> Е 1	<b>((</b> 2	<b>(1</b> 3	<b>11</b> 4	<b>11</b> 5	<b>((</b> 6	<b>(l</b> 7	<b>((</b> 8	<b>(1</b> 9	<b>(c</b> 10	<b>tt</b> 11			∎ X
<b> ?</b> F 1	<b>1</b> 2	<b>«</b> 3	<b>S</b> 4	<b>1</b> ( 5	6	(c 7	(c 8	<b>K</b> 9	<b>2C</b> 10	<b>)</b> (	<b>«</b> (12	<b>)(</b> 13	<b>) c</b> X <sub>1</sub> X <sub>2</sub>
<b>))</b> 14	<b>X</b> 15	<b>()</b> 16	(c 17	<b>(</b> 18	<b>(</b> 19	<b>R</b> 20							

**Figure 1.** Karyotypes of species based on spermatogonial metaphases. A *Drassyllus sur*,  $2n \stackrel{>}{_{-22}} = 22$ ,  $X_1 X_2 0$ **B** *Nomisia exornata*,  $2n \stackrel{>}{_{-22}} = 22$ ,  $X_1 X_2 0$  **C** *Nomisia orientalis*,  $2n \stackrel{>}{_{-22}} = 22$ ,  $X_1 X_2 0$  **D** *Sitticus caricis*,  $2n \stackrel{>}{_{-23}} = 28$ ,  $X_1 X_2 0$  **E** *Xysticus gallicus*,  $2n \stackrel{>}{_{-23}} = 23$ , X0 **F** *Pax islamita*  $2n \stackrel{>}{_{-24}} = 42$ ,  $X_1 X_2 0$ . Bar=10 µm.

The karyotype of *N. exornata* (C. L. Koch, 1839) (Fig. 1B)  $(2n \swarrow = 22, X_1X_20)$  was acrocentric. Autosome pairs decreased gradually in size from  $10.3\pm0.21\%$  to  $5.85\pm0.17\%$  of TCL. Relative length of X<sub>1</sub> and X<sub>2</sub> were  $7.46\pm0.13\%$  and  $6.65\pm0.08\%$  of TCL, respectively.

The autosomes of *N. orientalis* Dalmas, 1921 (Fig. 1C)  $(2n\Im = 22, X_1X_20)$  was acrocentric. RCL of autosome pairs were decreased gradually from 10.61±0.24% to 6.62±0.19% of TCL. The gonosomes  $X_1$  (7.91±0.12% of TCL) and  $X_2$  (6.10±0.07% of TCL) showed acrocentric morphology.

The sex chromosomes were positively heteropycnotic from leptotene to diakinesis in both *Nomisia* species studied. Plates consisted of 10 autosomal bivalents and two univalent sex chromosomes from pachytene to metaphase I (Fig. 2C–E). At meiotic anaphases, 10 chromosomes segregated to one pole and 12 chromosomes to another pole (Fig. 2F).

# Salticidae

The autosomes of *Sitticus caricis* (Westring, 1861)  $(2n_0^2=28, X_1X_20)$  were acrocentric. RCL decreased gradually from 8.47±0.42% to 5.04±0.16% of TCL (Fig. 1D). The sex

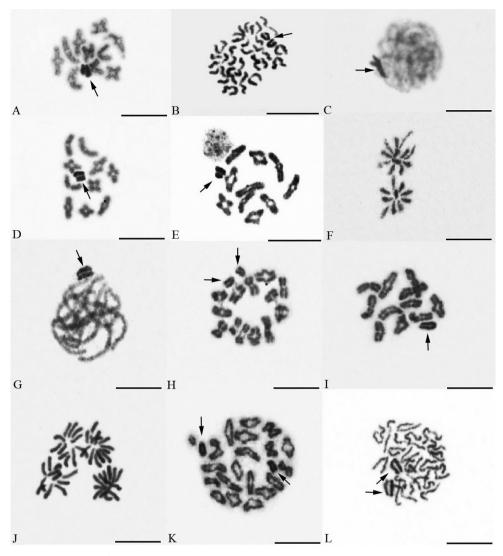


Figure 2. Meiosis of gnaphosid, salticid, thomisid and zodariid males. *Drassyllus sur* A diplotene B metaphase II, *Nomisia exornata* C early pachytene D diakinesis, *Nomisia orientalis* E diakinesis F part of anaphase II showing one plate with 10 chromosomes and another plate with 12 chromosomes, *Sitticus caricis* G pachytene H diplotene, *Xysticus gallicus* I diakinesis J anaphase II, *Pax islamita* K diakinesis L half of metaphase II (arrows indicate sex chromosomes). Bar=10 μm.

chromosomes X<sub>1</sub> (7.33 ±0.51% of TCL) and X<sub>2</sub> (6.72±0.38% of TCL) were medium sized in comparison with the autosomes.

Leptotene, zygotene, and pachytene nuclei included a positively heteropycnotic sex chromosome body that was located at the periphery of the nucleus (Fig. 2G). At late prophase I (i.e. diplotene and diakinesis), 13 autosomal bivalents and two univalent sex chromosomes were determined (Fig. 2H).

## Thomisidae

The chromosome set of *Xysticus gallicus* Simon, 1875  $(2n\partial = 23, X0)$  contained 11 acrocentric pairs and a small X chromosome (Fig. 1E). Autosome pairs decreased gradually in size from 10.28±0.62% to 6.46±0.39% of TCL. Relative length of X chromosome was 6.77±0.46% of TCL. This chromosome was longer than the smallest autosome pair.

From leptotene to diakinesis, X chromosome was formed by an intensively stained material. Diakinetic plates exhibit 11 autosomal bivalents (Fig. 2I). At metaphase II and anaphase II, X chromosome was isopycnotic with autosomes (Fig. 2J).

### Zodariidae

The karyotype of *Pax islamita* (Simon, 1873) consisted of acrocentric chromosomes; the diploid number was 42 (Fig. 1F). Autosome pairs exhibited a gradual decrease of relative lengths from  $6.42\pm0.58$  to  $3.31\pm0.24\%$  of TCL. This species showed  $X_1X_20$  sex chromosome system. The acrocentric gonosomes showed similar size. Their relative lengths were  $5.92\pm0.66\%$  and  $5.37\pm0.18\%$  of TCL, respectively.

From beginning of pachytene to metaphase I, plates consisted of 20 autosomal bivalents and two not associated sex chromosomes on the periphery of nucleus (Fig. 2K). Sex chromosomes were positively heteropycnotic during prophase and metaphase II. Metaphases II consisted of 20 or 22 chromosomes, respectively. Metaphases II with 22 chromosomes contained two X chromosomes (Fig. 2L).

## Discussion

Karyotypes of 771 spider species from 277 genera are known at present (Araújo et al. 2014). Diploid chromosome numbers of spiders range from 7 (Suzuki 1954) to 128 (Král et al. 2013). Entelegynae araneomorphs exhibit lower diploid numbers and mostly monoarmed chromosomes when compared with the predominantly high chromosome numbers and biarmed chromosomes of mygalomorphs (Kořínková and Král 2013). The sex chromosome system of most entelegynes is  $X_1X_2^0$  type. This system is supposed to be the ancestral form in spiders. It was found in more than 77% of spiders (Araújo et al. 2005).

So far, chromosome numbers have been established for 38 species of gnaphosid spiders. The majority of species (33 in a total) have  $2n^3 = 22$  including  $X_1X_20$  sex chromosome system (Araújo et al. 2014). Cytogenetics of *Drassyllus* is still not adequately explored. However, two karyotypes of *Drassyllus* have been published: *D. pumilus* (C.L. Koch, 1839)  $(2n^3 = 22, X_1X_20)$  (Kumbıçak et al. 2009) and *D. praeficus* (L. Koch, 1866)  $(2n^3 = 22, X_1X_20)$  (Kumbıçak et al. 2013). The same karyotype was found in *D. sur* (this study). Karyotypes of the *Nomisia* species analyzed show also the same karyotype with  $2n^3 = 22$  and  $X_1X_20$  sex chromosome system (Gorlova et al. 1997,

Kumbıçak et al. 2011, this study). *Drassyllus* and *Nomisia* belong to different subfamilies (Zelotinae and Gnaphosinae, respectively) (Ubick 2005, Seyyar et al. 2009). With one exception (*Urozelotes rusticus*, (L. Koch, 1872) Srivastava and Shukla 1986), all members of these subfamilies karyotyped so far presents  $2n^{-2}_{0}=22$ ,  $X_1X_20$  (Araújo et al. 2014), confirming the homogeneity of chromosome numbers, morphology and sex chromosomes systems in the family Gnaphosidae.

Male diploid numbers in salticids vary from 2n=14 in *Menemerus illigeri* (Audouin, 1826) (Gorlova et al. 1997) to 2n=28 in most salticids (109 in a total, Araújo et al. 2014). According to the previous studies, salticids exhibit considerable diversity of the sex chromosome systems (X0, X<sub>1</sub>X<sub>2</sub>0, X<sub>1</sub>X<sub>2</sub>X<sub>3</sub>0, and X<sub>1</sub>X<sub>2</sub>X<sub>3</sub>Y). 12 male karyotypes has been found in salticids, namely: 29, X<sub>1</sub>X<sub>2</sub>X<sub>3</sub>0; 28, X<sub>1</sub>X<sub>2</sub>0; 27, X0; 27, X<sub>1</sub>X<sub>2</sub>Y; 26, X<sub>1</sub>X<sub>2</sub>X<sub>3</sub>Y; 26, X<sub>1</sub>X<sub>2</sub>0; 25, X<sub>1</sub>X<sub>2</sub>Y; 25, X0; 23, X0; 22, X<sub>1</sub>X<sub>2</sub>0; 21, X0 and 14, X<sub>1</sub>X<sub>2</sub>0 (Araújo et al. 2014). Ancestral karyotype of salticids is probably formed by 28 chromosomes including X<sub>1</sub>X<sub>2</sub>0 system (Maddison and Leduc-Robert 2013). Our results showed  $2n \circ = 28$ , X<sub>1</sub>X<sub>2</sub>0 in *Sitticus caricis*. This chromosome number as well as the acrocentric chromosome morphology is the same as found in *Sitticus* species studied so far, namely *S. littoralis* (Hahn, 1832) (Suzuki 1954) and *S. terebratus* (Clerck, 1757) (Hackman 1948). *Sitticus* is the only genus belonging to Amycoida clade (Maddison and Hedin 2003) karyotyped so far. So, the current knowledge is not sufficient to explain the karyotype evolution of this clade, therefore new studies on the other amycoids are needed.

The male karyotype of *Xysticus gallicus* displays the general pattern described for most Thomisidae: a diploid chromosome number 23 and X0 sex chromosome system including acrocentric sex chromosome. All *Xysticus* species analyzed so far present this karyotype with exception of *X. triguttatus* Keyserling 1880 (Painter 1914). According to the phylogeny of Benjamin et al. (2008), *Xysticus* is sister group to *Coriarachne* Thorell, 1870, that also presents 2n=23, X0 (*C. fulvipes* Karsch, 1879; Suzuki 1952). There are two hypotheses explaining the origin of acrocentric X0 sex chromosome system in spiders. According to Datta and Chatterjee (1989, 1992) the acrocentric X chromosome can be derived by centric fusion of the X<sub>1</sub> and X<sub>2</sub> chromosomes, followed by pericentric inversions. Also, the acrocentric X chromosome could have originated from tandem fusion between acrocentric X<sub>1</sub> and X<sub>2</sub> chromosomes (Pekár and Král 2001).

Our study represented a diploid number of 42 acrocentric chromosomes and  $X_1X_20$  system in *Pax islamita*. This finding is compatible with the results reported by Král et al. (2011). However, these authors have also found heterozygotes for autosomal centric fusion  $(2n\Im = 41, X_1X_20)$  in addition to the standard individuals. Furthermore, they revealed different pattern of sex chromosome behaviour at male germline of this species. According to their results, the gonosomes were recognised as early as spermatogonial prophase and prometaphase due to their precocious condensation, positive heteropycnosis, and association. However, our data showed the sex chromosomes were indistinguishable at mitotic prophase and prometaphase from autosomes. It was possible to recognise them from autosomes at the beginning of meiotic prophase only due to positive heteropycnosis.

In conclusion, our study described the karyotype features of five araneomorph spiders for the first time and confirms some findings of Král et al. 2011 for *P. islamita*.

# Acknowledgements

We are very thankful to four anonymous referees for valuable suggestions and improvements of the manuscript. This study was carried out in the Department of Molecular Biology and Genetics, Faculty of Science and Art, Nevşehir Hacı Bektaş Veli University (Nevşehir, Turkey) and supported by Nevşehir Hacı Bektaş Veli University Directorate of Scientific Research Projects (NEUBAP13F10).

## References

- Araújo D, Cella DM, Brescovit AD (2005) Cytogenetic analysis of the neotropical spider *Nephilengys cruentata* (Araneomorphae, Tetragnathidae): standard staining, NORs, Cbands and base-specific fluorochromes. Brazilian Journal of Biology 65(2): 193–202. doi: 10.1590/S1519-69842005000200002
- Araújo D, Schneider MC, Paula-Neto E, Cella DM (2014) The spider cytogenetic database. Version 2.5. www.arthropodacytogenetics.bio.br/spiderdatabase [accessed on 23 February 2014]
- Benjamin SP, Dimitrov D, Gillespie RG, Hormiga G (2008) Family ties: molecular phylogeny of crab spiders (Araneae: Thomisidae). Cladistics 24(5): 708–722. doi: 10.1111/j.1096-0031.2008.00202.x
- Coddington JA, Levi HW (1991) Systematics and evolution of spiders (Araneae). Annual Review Ecology, Evolution and Systematics 22: 565–592. doi: 10.1146/annurev.es.22.110191.003025
- Datta SN, Chatterjee K (1989) Study of meiotic chromosomes of four hunting spiders of Northeastern India. Perspectives in Cytology and Genetics 6: 414–424.
- Datta SN, Chatterjee K (1992) Chromosomes and sex determination in three species of spinner spiders from Northeastern India. Cell and Chromosome Research 15(2): 64–69.
- Hackman W (1948) Chromosomenstudien an Araneen mit besonderer berücksichtigung der geschlechtschromosomen. Acta Zoologica Fennica 54: 1–101.
- Gorlova OYU, Gorlov IP, Nevo E, Logunov DV (1997) Cytogenetic studies on seventeen spider species from Israel. Bulletin of the British Arachnological Society 10(7): 49–252.
- Kořínková T, Král J (2013) Karyotypes, sex chromosomes, and meiotic division in spiders. In: Nenwing W (Ed) Spider Ecophysiology, 159–171.
- Král J, Kořínková T, Forman M, Krkavcová L (2011) Insights into the meiotic behavior and evolution of multiple sex chromosome systems in spiders. Cytogenetic and Genome Research 133(1): 43–66. doi: 10.1159/000323497
- Král J, Kořínková T, Krkavcová L, Musilová J, Ávila Herrera IM, Forman M, Vitkova M, Haddad CR, Hedin M, Henriques S, Palacios Vargas JG (2013) Evolution of the karyotype, sex chromosome systems, and meiosis in mygalomorph spiders (Araneae: Mygalomorphae). Biological Journal of the Linnean Society 109(2): 377–408. doi: 10.1111/bij.12056
- Kumbıçak Z, Ergene S, Saygıdeğer S (2009) Chromosomal data on six araneomorph spiders belonging to the families Lycosidae and Gnaphosidae (Araneae: Araneomorphae). Zoology in the Middle East 48(1): 89–96. doi: 10.1080/09397140.2009.10638371

- Kumbıçak Z (2010) Investigations on karyotypes and sex chromosome systems of some spiders in Turkey. Ph.D. Dissertation, Science Institute, Gaziantep University, 155 pp. [In Turkish].
- Kumbıçak Z, Ergene S, Karataş A, Kumbıçak Ü (2011) Cytogenetic studies on five species of spiders from Turkey (Araneae: Gnaphosidae, Lycosidae). Journal of Arachnology 39(3): 490–494. doi: 10.1636/B11-06.1
- Kumbıçak Z, Karataş A, Kumbıçak Ü, Seyyar O (2013) Karyological data and meiosis of *Drassyllus praeficus* (L. Koch, 1866) (Gnaphosidae) and *Thanatus imbecillus* (L. Koch, 1878) (Philodromidae) from Turkey. Turkish Journal of Zoology 37(2): 200–204.
- Levan A, Fredga K, Sandberg AA (1964) Nomenclature for centromeric position on chromosomes. Hereditas 52(2): 201–220. doi: 10.1111/j.1601-5223.1964.tb01953.x
- Maddison WP, Hedin MC (2003) Jumping spider phylogeny (Araneae:Salticidae). Invertebrate Systematics 17: 529–549. doi: 10.1071/IS02044
- Maddison WP, Leduc-Robert G (2013) Multiple origins of sex chromosome fusions correlated with chiasma localization in *Habronattus* jumping spiders (Araneae: Salticidae). Evolution 67(8): 2258–2272. doi: 10.1111/evo.12109
- Painter TS (1914) Spermatogenesis in spiders. Zoologische Jahrbuecher Abteilung fuer Anatomie und Ontogenie der Tiere 38: 509–576.
- Platnick NI (2014) The World Spider Catalog. Version 14.5. American Museum of Natural History. http://research.amnh.org/iz/spiders/catalog [accessed on 26 February 2014]
- Pekár S, Král J (2001) A comparative study of the biology and karyotypes of two central European zodariid spiders (Araneae, Zodariidae). Journal of Arachnology 29(3): 345–353. doi: 10.1636/0161-8202(2001)029[0345:ACSOTB]2.0.CO;2
- Seyyar O, Ayyıldız N, Topçu A (2009) Description of a new species of the genus *Nomisia* Dalmas, 1921 (Araneae: Gnaphosidae) from Turkey with some faunistical remarks. Zootaxa 2006: 62–68.
- Srivastava MDL, Shukla S (1986) Chromosome number and sex-determining mechanism in forty-seven species of Indian spiders. Chromosome Information Service, 41: 23–26.
- Suzuki S (1952) Cytological studies in spiders II. Chromosomal investigation in twenty two species of spiders belonging to the four families, Clubionidae, Sparassidae, Thomisidae and Oxyopidae, which constitute Clubionoidea, with special reference to sex chromosomes. Journal of Science of the Hiroshima University, Series B, Division 1, 13: 1–52.
- Suzuki S (1954) Cytological studies in spiders. III. Studies on the chromosomes of fifty-seven species of spiders belonging to seventeen families, with general considerations on chromosomal evolution. Journal of science of the Hiroshima University, Series B. Division 1, 15(2): 23–136.
- Traut W (1976) Pachytene mapping in the female silkworm *Bombyx mori* L. (Lepidoptera). Chromosoma 58(3): 275–284. doi: 10.1007/BF00292094
- Ubick D (2005) Gnaphosidae In: The Spider of North America: An Identification Manual. Ed: America Arachnological Society, 69–74.