

Chromosome numbers and DNA content in some species of *Mecardonia* (Gratiolae, Plantaginaceae)

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Abstract

Cytogenetic characterization and determination of DNA content by flow cytometry of five species of *Mecardonia* Ruiz et Pavon, 1798 (Gratiolae, Plantaginaceae) was performed. This is the first study of nuclear DNA content carried out in the genus. Mitotic analysis revealed a base chromosome number $x = 11$ for all entities and different ploidy levels, ranging from diploid ($2n = 2x = 22$) to hexaploid ($2n = 6x = 66$). The results include the first report of the chromosome numbers for *M. flagellaris* (Chamisso & Schlechtendal, 1827) ($2n = 22$), *M. grandiflora* (Benth) Pennell, 1946 ($2n = 22$), *M. kamogawae* Greppi & Hagiwara, 2011 ($2n = 66$), and *Mecardonia* sp. ($2n = 44$). The three ploidy levels here reported suggest that polyploidy is common in *Mecardonia* and appear to be an important factor in the evolution of this genus. The $2C$ - and $1Cx$ -values were also estimated in all the species. The $2C$ -values ranged from 1.91 to 5.29 pg. The $1Cx$ -values ranged from 0.88 to 1.03 pg. The general tendency indicated a decrease in the $1Cx$ -value with increasing ploidy level. The significance of the results is discussed in relation to taxonomy of the genus.

Keywords

Gratiolae, chromosome number, DNA content, flow cytometry, polyploidy

Introduction

Mecardonia Ruiz & Pavon, 1798 belongs to the tribe Gratiolae (Plantaginaceae) and is distributed across the America, reaching its southernmost distribution in Argentina. The species are erect or creeping herbs, annual or perennial, much branched, mostly glabrous, sometimes blackening on drying, gland dotted, and yellow and white flowers (D'Arcy 1979). Since the description of the genus (Ruiz and Pavon 1794), there have been a few problems in establishing its generic and infrageneric circumscription. Rossow (1987) considered only 10 species, which had been previously placed in *Bacopa* Aublet, 1775 under subgenus *Mecardonia* (Descole and Borsini 1954). More recently, Souza (1997) and Souza and Giuletti (2009) carried out some taxonomic modifications to Rossow's classification. The demarcation of the genus is variable depending on the author consulted. Following Rossow's classification, the genus includes five species growing in Argentina: *M. flagellaris* (Chamisso & Schlechtendal, 1827), *M. grandiflora* (Bentham) Pennell, 1946, *M. procumbens* Small, 1903, *M. serpylloides* (Chamisso & Schlechtendal, 1891) and *M. tenella* (Chamisso & Schlechtendal, 1891). Recently, *M. kamogawae* Greppi & Hagiwara, 2011 was described by as an endemic species of Corrientes Province (Argentina).

Mecardonia has ornamental value because some cultivars developed from native species from Northern Argentina were recently introduced in the trade of ornamental plants (Greppi 2012). Therefore, researches on genetic improvement are carried out in this genus.

Cytological and cytogenetic studies have proved useful data for taxonomic and evolutionary analyses, which are widely used in processes of conventional or biotechnological genetic improvement (Poggio et al. 2004). Characters such as chromosome number, morphology, and meiotic behavior, as well as nuclear DNA content, have been used as taxonomic markers helping to circumscribe taxa and infer their relationships (Kron et al. 2007, Guerra 2008, Loureiro et al. 2010, Castro et al. 2012). At present, only two species of *Mecardonia* have been evaluated cytologically. Lewis et al. (1962) reported $2n = 42 \pm 2$ for a Northamerican species *M. acuminata* (Walter, 1891) Small, 1903. Kaul (1969) determined $2n = 2x = 22$ for *M. procumbens* Small, 1903 (as *M. dianthera* (Swartz 1900, Pennell 1946). Therefore, to increase the knowledge of *Mecardonia*, other species were cytologically analyzed in this study.

Nuclear DNA content, understood as genome size, is very variable across angiosperm, and has been revealed as an important character in biodiversity. In *Mecardonia* species there are no reported measurements of DNA content, but genome size variation has been explored in some genera of Plantaginaceae. DNA C-values are currently available for 204 species belonging to 18 genera of this family, including *Callitriche* Linnaeus, 1753, *Penstemon* Schmidel, 1762, *Plantago* Linnaeus, 1753 and *Veronica* Linnaeus, 1753 with a range of variation of 0.32–4.63 pg (Albach and Greilhuber 2004, Broderick et al. 2011, Bennett and Leitch 2012, Wong and Murray 2012, Prančl et al. 2014, Meudt et al. 2015). Herein we used flow cytometry to estimate the nuclear DNA content in five species of *Mecardonia* for the first time. Additionally, we report original chromosome numbers of some of them. The results are discussed in relation to the taxonomy and evolution of the genus.

Materials and methods

We examined six populations from five species of *Mecardonia* collected in Argentina. Information about the studied material and the voucher specimens is provided in Table 1. Vouchers are deposited at the herbarium BAB of the Instituto Nacional de Tecnología Agropecuaria (INTA), Buenos Aires, Argentina.

Mitosis analysis

Mitotic chromosome preparations were made from root meristems obtained from rooted stems. The roots were pretreated for about 4 h in 0.002 M 8-hydroxyquinoline solution at room temperature, fixed in 5:1 absolute alcohol/lactic acid, and then stained using Feulgen's technique. Permanent microscope slides were prepared by mounting in Euparal. In all samples at least 20 counts of 7–10 individuals were made to verify the observations.

Permanent microscope slides were examined and photographed using Zeiss Axioplan microscope Carl Zeiss with digital camera Canon Power Shot A 640.

Nuclear DNA measurements

DNA content (in picograms) was estimated by flow cytometry using fresh young leaves. The measurements were calculated from three replicates per individuals. In total we analyzed three individuals per species. The leaves of *Zea mays* Linnaeus, 1753 cv. 'CE-777' ($2C = 5.43$ pg., Doležel et al. 1998) were used as internal standard for almost all entities. While, *Hordeum vulgare* Linnaeus, 1753 cv. 'New Golden' ($2C = 10.4$ pg., Bennett

Table 1. *Mecardonia* species analyzed in this study, with their respective chromosome numbers ($2n$), locations, and voucher specimens.

	Species	$2n$	Location, voucher specimen
*	<i>M. flagellaris</i> (Cham. & Schldt.) Rossow	$2n = 2x = 22$	Argentina. Entre Ríos, Dept. Federación, in front of complejo turístico Irupé. Greppi et al. 1411 (BAB).
		$2n = 2x = 22$	Argentina. Entre Ríos, Dept. Federación, complejo turístico Irupé Greppi et al. 1190 (BAB).
*	<i>M. grandiflora</i> (Benth.) Pennell	$2n = 2x = 22$	Argentina. Misiones, Dept. Guaraní, Ayo. Pepirí Miní o Yabotí. Greppi et al. 1189 (BAB).
*	<i>M. kamogawae</i> Greppi & Hagiwara	$2n = 6x = 66$	Argentina. Corrientes, Dept. Paso de los Libres, Paso de los Libres to national route 14, Greppi et al. 1081 (BAB).
	<i>M. procumbens</i> (Mill.) Small	$2n = 2x = 22$	Argentina. Córdoba. Dept. Unión, Monte Leña, national route 9, km 491, Greppi 681 (BAB).
*	<i>Mecardonia</i> sp. n.	$2n = 4x = 44$	Argentina. Corrientes. Dept. Empedrado. Greppi and Hagiwara 1410 (BAB)

* First chromosome count.

and Leitch 2005) was used as the standard of hexaploid species. The selection of these internal standards was made since they are the common standards used in the laboratory where the flow cytometer is situated (Instituto de Floricultura, INTA Castelar, Buenos Aires, Argentina). To release nuclei from the cells, 0.5 cm² of leaf tissue of *Mecardonia* was chopped together with 0.5 cm² of leaf tissue of the internal standard in 0.5 ml buffer (High resolution DNA kit, Partec GmbH, Münster, Germany). Subsequently, 5 U ml⁻¹ of RNase were added and incubated for 2–5 min at room temperature. The suspension was filtered through a 30 µm nylon mesh. After this period, 1.5 ml of staining solution containing 1 µg µl⁻¹ propidium iodide was added. Within 1 h of staining, measurements were performed with a CyFlow Ploidy Analyzer, Partec cytometer (green laser 532 nm, 30 mW). About 10,000 nuclei were measured for each sample.

The absolute value of DNA content (2C) of each sample was calculated by the formula: (\bar{X} peak of sample × G1 DNA content (2C) of the standard)/ \bar{X} G1 peak of the standard (Doležel and Bartos 2005).

The monoploid genome size (1Cx) was calculated dividing the 2C-value by the ploidy level (Greilhuber et al. 2005).

Data analysis

The mean, standard deviation and the coefficient of variation of 2C-value were calculated for each species from three different individuals. Differences in 1Cx-value between species were tested by one-way analysis of variance (ANOVA) at a significance level of 5% ($\alpha = 0.05$). The Tukey 5% post hoc test was used to test differences between each pair of species.

Pearson correlation coefficient was calculated to test whether the 2C- and 1Cx-values were related to chromosome number. Scatter plot was performed to evaluate the relationship between the 1Cx-values and the chromosome numbers (2n) of species. All statistical analyses were performed using the InfoStat software version 2013 (Di Rienzo et al. 2013).

Results

The chromosome numbers of six populations belonging to five species of *Mecardonia* were determined. The analyzed species and their chromosome numbers are given in Table 1. Four species were analyzed for the first time. The chromosome number observed in the remaining taxon is in agreement with previous studies. All species analysed shared the same base chromosome number ($x = 11$), and chromosome numbers ranged between $2n = 22$ to $2n = 66$. Of these, only three species were diploids: *M. flagellaris* (Fig. 1 A), *M. grandiflora* (Fig. 1 B) and *M. procumbens*. The remaining species were polyploids, *Mecardonia* sp. (Fig. 1 C) was tetraploid with $2n = 44$ and *M. kamogawae* was hexaploid with $2n = 66$ (Fig. 1 D).

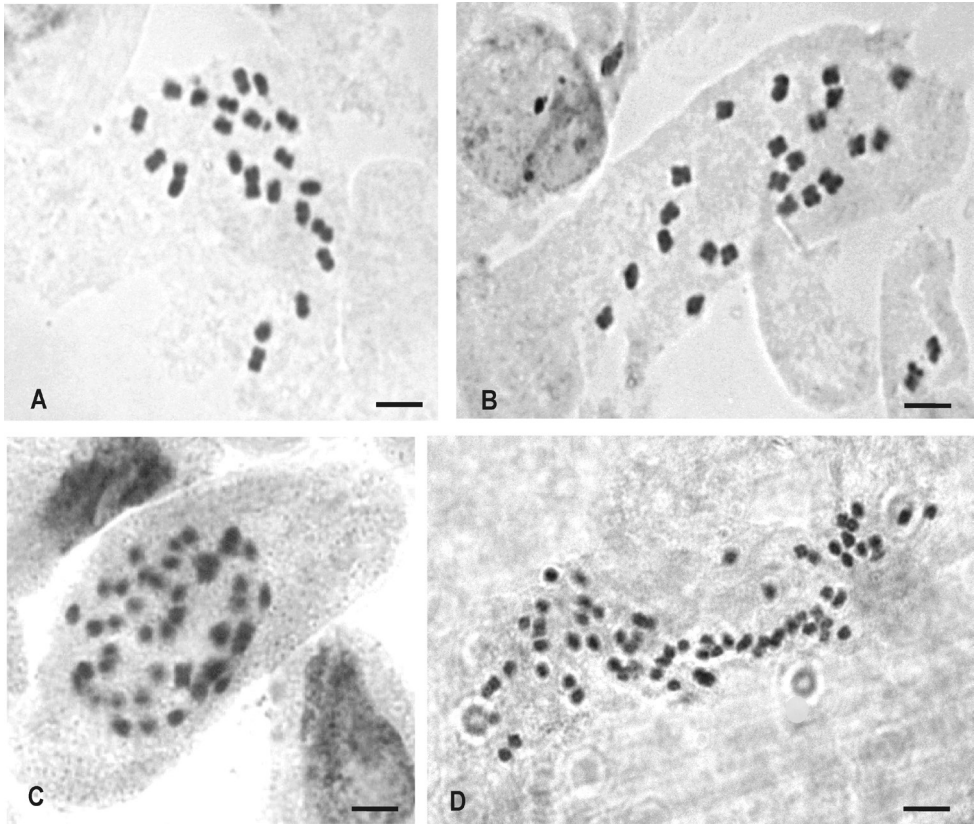


Figure 1. Somatic chromosomes of *Mecardonia* species. **A** *M. flagellaris*: $2n = 2x = 22$ **B** *M. grandiflora*: $2n = 2x = 22$ **C** *Mecardonia* sp. n.: $2n = 4x = 44$ **D** *M. kamogawae*: $2n = 6x = 66$. Bar = 5 μm .

Genomic DNA content

The DNA amounts determined for five species of *Mecardonia* are shown in Table 2. The flow cytometric measurements of all species and the standards resulted in well defined and sharp peaks. In all cases, the coefficients of variation were lower than 5% (Table 2), supporting the reliability of the flow cytometric assessments.

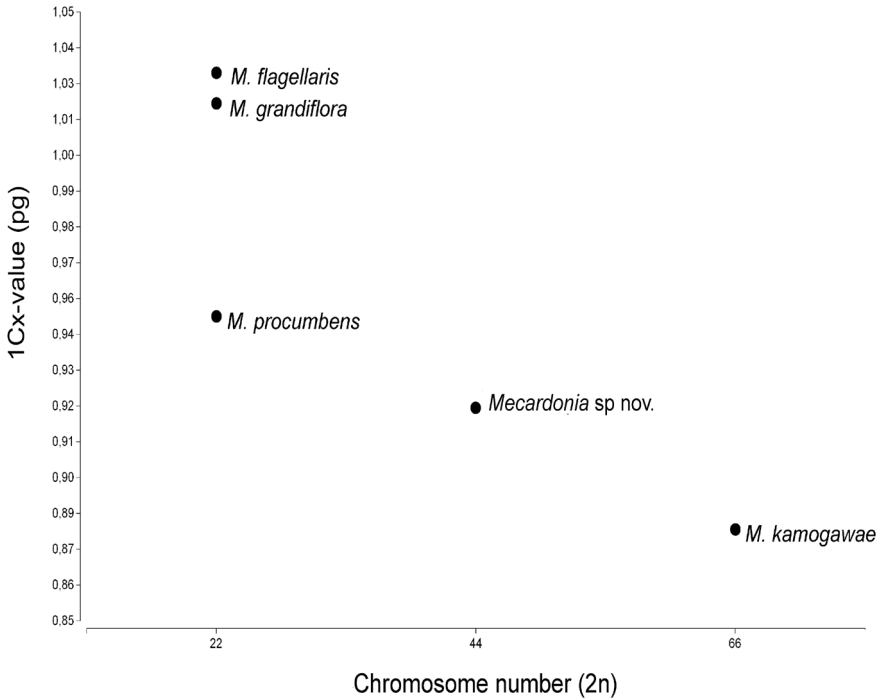
The 2C-values of the species here analyzed varied from 1.91 pg in *M. procumbens* (2x) to 5.29 pg in *M. kamogawae* (6x). The 2C-values were strongly and significantly correlated with chromosome number ($r = 0.99$; $P = < 0.0001$).

The 1Cx-values, which indicated the DNA content per genome, ranging from 1Cx = 0.88 pg in *M. kamogawae* to 1Cx = 1.03 pg in *M. flagellaris* (Table 2). The ANOVA showed significant differences for 1Cx-values ($F = 357.52$; $P = < 0.0001$) between the species. The correlation between values of 1Cx and chromosome number was negative and significant ($r = -0.86$; $P = < 0.0001$, Fig. 2).

Table 2. Chromosome number (2n), ploidy level, 2C-value (pg), CV (coefficient of variation), 1Cx-value (pg) of the *Mecardonia* species analyzed.

Species	Chromosome number (2n)	Ploidy level	2C (pg)	CV	1Cx (pg)
<i>M. flagellaris</i>	22	2x	2.06 ± 0.16	0.029	1.03 ^d
<i>M. grandiflora</i>	22	2x	2.05 ± 0.08	0.010	1.02 ^d
<i>M. procumbens</i>	22	2x	1.91 ± 0.06	0.012	0.95 ^c
<i>Mecardonia</i> sp. n.	44	4x	3.71 ± 0.05	0.053	0.92 ^b
<i>M. kamogawae</i>	66	6x	5.29 ± 0.10	0.061	0.88 ^a
ANOVA					(F=357.52; P= <0.0001)

For ANOVA results, different lower-case letters indicate significant differences among species for mean values of each parameter at 5% level using Tukey's test.

**Figure 2.** Scatter plot between 1Cx-value and chromosome number (2n).

Discussion

The chromosome number $2n = 22$ found in *M. procumbens*, is consistent with the chromosome counts recorded in a previous cytological study (Kaul 1969). Sinha (1987) reported B chromosomes for this taxon; however, the populations here analyzed did not show these accessory chromosomes. Chromosome counts for *M. flagellaris* ($2n = 22$), *M. grandiflora* ($2n = 22$), *M. kamogawae* ($2n = 66$) and *Mecardonia* sp. ($2n = 44$) are

described here for the first time. Our results showed diploid and polyploid species for the genus. Polyploidization has long been recognized as an important process in plant evolution (Otto and Whitton 2000, Soltis et al. 2004). In Plantaginaceae, polyploidy is a common phenomenon occurring in many genera, such as *Antirrhinum* Linnaeus, 1753, *Chaenorhinum* (DC.) Reichenbach, 1828, *Cymbalaria* Hill, 1756, *Chelone* Linnaeus, 1753, *Digitalis* Linnaeus, 1753, *Linaria* Miller, 1754, *Plantago*, *Nuttallanthus* D.A. Sutton, 1988, *Stemodia* Linnaeus, 1759, *Veronica* (Hair 1966, Subramanian and Pondmudi 1987, Sosa and Seijo 2002, Sosa et al. 2009, 2011, Wolfe et al. 2002, Vargas et al. 2004, Murray et al. 2010, Castro et al. 2012, Wong and Murray 2012, Ranjbar and Nouri 2015). Our results evidenced the presence of multiple cytotypes in *Mecardonia*, hence suggesting polyploidy as a key driver of the evolution of the genus. The present analysis, in addition to previous chromosome number reports, revealed that the genus have exclusively the basic chromosome number of $x = 11$.

The interest on the study of genome size increased in the last decade. These studies focused on the use of genome size as a taxonomic marker (Castro et al. 2012, Angulo and Dematteis 2013, Galdeano et al. 2016) and on finding correlations between ecological and environmental variables and this character (Chalup et al. 2014, Vega and Dematteis 2015). However, there are still many families and genera being neglected, including *Mecardonia*, for which the present study is the first analysis of genome size for the genus. The estimates 2C- and 1C-values calculated for the species in this study are within ranges of variation found in Angiosperms and Plantaginaceae (Leitch and Bennett 2004, Meudt et al. 2015). Based on the available genome size data, *Mecardonia* falls into the categories “very small” genomes (2C = <2.8 pg) to “small” genomes (2C = <7.0 pg) according to values reported by Leitch et al. (1998) and Soltis et al. (2004).

The 2C-values of *Mecardonia* species revealed a positive and significant correlation with chromosome number ($r = 0.99$, $P = < 0.0001$). Therefore, in the genus there is a trend for increasing 2C-value with increasing ploidy level. On the other hand, the variation of 1Cx-values is negative and significantly ($r = -0.86$; $P = < 0.0001$) correlated with chromosome number. Consequently, the values of 1Cx of the species decrease in inverse proportional to the ploidy level. Our data reflect that both polyploids (tetraploid 1Cx = 0.92 pg and hexaploid 1Cx = 0.88 pg) have lesser values of monoploid genome size than diploid species (mean of Cx = 1.00 pg). Many polyploid angiosperms undergo *genome downsizing* and so have smaller average genome sizes than their diploid relatives (Leitch and Bennett 2004, Leitch et al. 2008) and *Mecardonia* seems not to be an exception. Several studies have indicated that during polyploidization different balancing processes at genomic level occur which may promote variation in nuclear DNA content. These changes point towards a possible need for harmonization of genome and removal of some unnecessary genomic redundancies (Petrov 2001; Bennetzen et al. 2005, Pellicer et al. 2010). Mechanisms leading to a decrease in genome size in polyploids may include non-random elimination of chromosome- and genome-specific sequences (Ozkan et al. 2003, Shaked et al. 2001), illegitimate crossing over (Devos et al. 2002) or unbalanced deletion–insertion rates (Petrov 2001, 2002). Counterbalancing mechanisms are probably also involved to reduce the genetic and structural instabilities that accompany DNA loss (Pellicer et al. 2010).

Recently, Meudt et al. (2015) established a relationship between the genome downsizing with diversification in polyploid lineages of *Veronica* (Plantaginaceae), but they do not know how general this pattern might be or what causes it. Several hypotheses have been proposed to explain this relationship. Kraaijeveld (2010) suggested that organisms with small genomes have more stably inherited mutations, or a nucleotypic effect, in which organisms with small genomes and shorter genes have a general advantage as a result of faster replication and transcription. The genome size changes in *Mecardonia* are probably related with the diversification of the species. Further studies comparing this genus with the closest extant relative to determine what aspect of genome downsizing facilitate diversification are needed.

Taxonomic implications

The genus *Mecardonia* is currently under revision and some closely related species with intermediate morphological characteristics were found. It has been well documented in many plants that chromosome numbers and genome size can be used as extra taxonomic characters for discriminating between closely related taxa, helping to clarify the taxonomy of some species in problematic genus (Guerra 2008, Castro et al. 2012, Sosa and Dematteis 2014). For example, *Mecardonia* sp. is closely related to *M. flagellaris*. A detailed morphological analysis along with chromosome number here reported showed that it should be considered as different species. *Mecardonia* sp. is tetraploid with $2n = 44$, while *M. flagellaris* is diploid with $2n = 22$. Thus, both species differ in chromosome number and morphological features, such as aspect of plant, leaf shape, and trichome types of corolla. In addition, the new species has more restricted distribution to North of Argentina. However, *M. flagellaris* is expanding from Mato Grosso do Sul (Brazil) to Chubut (Argentina), arrived to Chile, Paraguay and Uruguay.

Another case is *M. kamogawae* that is morphologically related to *M. procumbens* from which it differs in the life-form, root types, leaf texture, and size of bracteoles and pedicels. Regarding chromosome number, *M. kamogawae* is hexaploid with $2n = 66$, while *M. procumbens* is diploid with $2n = 22$. Therefore, both species can be distinguished by morphological features, as well as by the chromosome number.

Mecardonia procumbens and *M. flagellaris* were diploids with $2n = 22$. Although the chromosome number does not distinguish both species, differences in 2C-values were observed. *Mecardonia flagellaris* had higher value ($2C = 2.06$ pg.) than *Mecardonia procumbens* ($2C = 1.91$ pg.). D'Arcy (1979) and Souza (1997) placed *M. flagellaris* under the synonymy of *M. procumbens* by having similar morphological characteristics. We considered, however, that both species are morphologically distinct. *Mecardonia procumbens* differs by aspect of the plant, shape and length of leaf, and calyx pieces. Also, *M. procumbens* has a wider distribution as it extends from the South of the United States of America to the Argentine Northwest. However, *M. flagellaris* grow in Paraguay, South of Brasil, Uruguay and Northeast of Argentina.

Conclusion

The results of this study suggest that chromosome number is useful in distinguishing species of *Mecardonia*. The different ploidy levels of the taxa showed the importance of polyploidy in the evolution of the genus. The results here obtained combined with those reported previously confirm that the *Mecardonia* genus has basic number $x = 11$.

Regarding to the variation of genome size, decreases in DNA content have occurred during the evolution of genome size in the *Mecardonia* species.

Our results showed that differences in morphological features along with chromosome numbers and DNA content values support Rossow's criterion (1987).

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References

- Albach DC, Greilhuber J (2004) Genome size variation and evolution in *Veronica*. *Annals of Botany* 94: 897–911. <https://doi.org/10.1093/aob/mch219>
- Angulo MB, Dematteis M (2013) Nuclear DNA content in some species of *Lessingianthus* (Vernonieae, Asteraceae) by flow cytometry. *Journal of Plant Research* 126: 461–468. <https://doi.org/10.1007/s10265-012-0539-x>
- Aublet JBCF (1775) *Histoire des Plantes de la Guiane Française* 1. Pierre – François Didot, London & Paris, 128–130.
- Bennett MD, Leitch IJ (2005) Nuclear DNA amounts in angiosperms: progress, problems and prospects. *Annals of Botany* 95: 45–90. <https://doi.org/10.1093/aob/mci003>
- Bennett MD, Leitch IJ (2012) Plant DNA C-values database (release 6.0, Dec. 2012). <http://www.kew.org/cvalues> [accessed 27 October 2016]
- Bennetzen JL, Ma J, Devos KM (2005) Mechanisms of recent genome size variation in flowering plants. *Annals of Botany* 95(1): 127–132. <https://doi.org/10.1093/aob/mci008>
- Broderick SR, Stevens MR, Geary B, Love SL, Jellen EN, Dockter RB, Dayley SL, Lindgren DT (2011) A survey of *Penstemon*'s genome size. *Genome* 54(2): 160–173. doi: 10.1139/G10-106
- Castro M, Castro S, Loureiro J (2012) Genome size variation and incidence of polyploidy in Scrophulariaceae sensu lato from the Iberian Peninsula. *AoB PLANTS* 2012: pls037. <https://doi.org/10.1093/aobpla/pls037>

- Chalup L, Grabile M, Neffa VS, Seijo G (2014) DNA content in South American endemic species of *Lathyrus*. Journal of Plant Research 127(4): 469–480. <https://doi.org/10.1007/s10265-014-0637-z>
- D'Arcy WG (1979) Scrophulariaceae. In: Woodson RR, Schery RW (Eds) Flora de Panamá. Parte IX. Annales of Missouri Botanical Garden 66: 173–274.
- Devos KM, Brown JK, Bennetzen JL (2002) Genome size reduction through illegitimate recombination counteracts genome expansion in *Arabidopsis*. Genome Research 12(7): 1075–1079. <https://doi.org/10.1101/gr.132102>
- Descole HR, Borsini OH (1954) Scrophulariaceae-Anthirinoideae. In: Descole HR (Ed.) Genera Species Plantarum Argentinarum 5: 131–151.
- Di Rienzo JA, Casanoves F, Balzarini MG, Gonzalez L, Tablada M (2013) InfoStat versión 2013. Grupo InfoStat, FCA, Universidad Nacional de Córdoba. <http://www.infostat.com.ar>
- Doležel J, Bartos J (2005) Plant DNA flow cytometry and estimation of nuclear genome size. Annals of Botany 95: 99–110. <https://doi.org/10.1093/aob/mci005>
- Doležel J, Greilhuber J, Lucretti S, Meister A, Lysák MA, Nardi L, Obermayer R (1998) Plant genome size estimation by flow cytometry: inter-laboratory comparison. Annals of Botany 82: 17–26.
- Galdeano F, Urbani MH, Sartor ME, Honfi AI, Espinoza F, Quarín CL (2016) Relative DNA content in diploid, polyploid, and multiploid species of *Paspalum* (Poaceae) with relation to reproductive mode and taxonomy. Journal of Plant Research 129(4): 697–710. <https://doi.org/10.1007/s10265-016-0813-4>
- Greilhuber J, Doležel J, Lysák MA, Bennett MD (2005) The origin, evolution and proposed stabilization of the terms 'genome size' and 'C-value' to describe nuclear DNA contents. Annals of Botany 95(1): 255–260. <https://doi.org/10.1093/aob/mci019>
- Greppi JA (2012) Manual de Cultivos de *Mecardonia*: variedades comerciales de *Mecardonia*: Poty Amarilla INTA y Guaraní Amarilla INTA. Instituto de Floricultura, 9 pp.
- Greppi JA, Hagiwara JC (2011) Una nueva especie de *Mecardonia* (Plantaginaceae). Darwiniana 49: 43–46.
- Guerra M (2008) Chromosome numbers in plant cytogenetics: concepts and implications. Cytogenetic Genome Research 120: 339–350. <https://doi.org/10.1159/000121083>
- Hair JB (1966) Biosystematics of the New Zealand flora, 1945–1964. New Zealand Journal of Botany 4: 559–595. <https://doi.org/10.1080/0028825X.1966.10430184>
- Hill J (1756) The British Herbal: An History of Plants and Trees, Natives Britain, Cultivated for Use, or Raised for Beauty. T. Osborne and J. Shipton, London, 113–114. <http://biodiversitylibrary.org/page/34898152>
- Kaul MLH (1969) Cytogenetical studies on ecological races of *Mecardonia dianthera* (SW) Pennell I. Cytology, floral-biology and pollination mechanism. Cytologia 34: 169–177. <http://dx.doi.org/10.1508/cytologia.34.169>
- Kraaijeveld K (2010) Genome size and species diversification. Evolutionary Biology 37(4): 227–233. <https://doi.org/10.1007/s11692-010-9093-4>
- Kron P, Suda J, Husband BC (2007) Applications of flow cytometry to evolutionary and population biology. Annual Review of Ecology, Evolution, and Systematics, 847–876. <https://doi.org/10.1146/annurev.ecolsys.38.091206.095504>
- Leitch IJ, Bennett MD (2004) Genome downsizing in polyploid plants. Biological Journal of Linn. Soc. 82: 651–663. <https://doi.org/10.1111/j.1095-8312.2004.00349.x>

- Leitch IJ, Chase MW, Bennett MD (1998) Phylogenetic analysis of DNA C-values provides evidence for a small ancestral genome size in flowering plants. *Annals of Botany* 82(suppl 1): 85–94. <https://doi.org/10.1006/anbo.1998.0783>
- Leitch IJ, Hanson L, Lim KY, Kovarik A, Chase MW, Clarkson JJ, Leitch AR (2008) The ups and downs of genome size evolution in polyploid species of *Nicotiana* (Solanaceae). *Annals of Botany* 101(6): 805–814. <https://doi.org/10.1093/aob/mcm326>
- Lewis WH, Stripling HL, Ross RG (1962) Chromosome numbers for some angiosperms of the southern United States and Mexico. *Rhodora* 64(758): 147–161.
- Linnaeus C (1753) *Species Plantarum*, Impresis Laurentii Salvii.
- Linnaeus C (1759) *Systema Naturae*, Tenth Edition. Impresis Laurentii Salvii.
- Loureiro J, Trávníček P, Rauchová J, Urfus T, Vit P, Štech M, Castro S, Suda J (2010) The use of flow cytometry in the biosystematics, ecology and population biology of homoploid plants. *Preslia* 82(1): 3–21.
- Meudt HM, Rojas-Andrés BM, Prebble JM, Low E, Garnock-Jones PJ, Albach DC (2015) Is genome downsizing associated with diversification in polyploid lineages of *Veronica*? *Botanical Journal of the Linnean Society* 178(2): 243–266. <https://doi.org/10.1111/boj.12276>
- Miller P (1754) *The Gardeners Dictionary Abridged* (4th edn). London.
- Murray BG, Meudt HM, Tay ML, Garnock-Jones PJ (2010) New chromosome counts in New Zealand species of *Plantago* (Plantaginaceae). *New Zealand Journal of Botany* 48: 197–204. <https://doi.org/10.1080/0028825x.2010.515598>
- Otto SP, Whitton J (2000) Polyploid incidence and Evolution. *Annual of Review Genetic* 34: 401–437. <https://doi.org/10.1146/annurev.genet.34.1.401>
- Ozkan H, Tuna M, Arumuganathan K (2003) Nonadditive changes in genome size during allopolyploidization in the wheat (*Aegilops-Triticum*) group. *Journal of Heredity* 94(3): 260–264. <https://doi.org/10.1093/jhered/esg053>
- Pellicer J, Garcia S, Canela MA, Garnatje T, Korobkov AA, Twibell JD, Vallès J (2010) Genome size dynamics in *Artemisia* L. (Asteraceae): following the track of polyploidy. *Plant Biology* 12(5): 820–830. <https://doi.org/10.1111/j.1438-8677.2009.00268.x>
- Pennell FW (1946) Reconsideration of the Bacopa-Herpestis problem of the Scrophulariaceae. *Proceedings of the Academy of Natural Sciences of Philadelphia* 98: 83–98.
- Petrov DA (2001) Evolution of genome size: new approaches to an old problem. *Trends of Genetic* 17: 23–28. [http://dx.doi.org/10.1016/S0168-9525\(00\)02157-0](http://dx.doi.org/10.1016/S0168-9525(00)02157-0)
- Petrov DA (2002) Mutational equilibrium model of genome size evolution. *Theoretical Population Biology* 61(4): 531–544. <https://doi.org/10.1006/tpbi.2002.1605>
- Poggio L, González G, Ferrari MR, García AM, Wulff A, Greizerstein E, Tomas P, Schrauf G (2004) Aportes de la Citogenética al Estudio de Genomas Vegetales (Parte V) – Ejemplos de Aplicaciones de la Biotecnología Vegetal (Capítulo 1). In: Levitus G, Echenique V, Rubinstein C, Hopp E, Mroginski L (Eds) *Biotecnología y Mejoramiento Vegetal II*. Consejo Argentino para la Información y el Desarrollo de la Biotecnología, 379–387.
- Pranč J, Kaplan Z, Trávníček P, Jarolímová V (2014) Genome size as a key to evolutionary complex aquatic plants: polyploidy and hybridization in *Callitriche* (Plantaginaceae). *PLoS ONE* 9(9). <http://dx.doi.org/10.1371/journal.pone.0105997>
- Ranjbar M, Nouri S (2015) Biogeography of the Genus *Linaria* (Plantaginaceae) Based on Chromosome Number Data. *Journal of Cell and Molecular Research* 7: 115–132.

- Reichenbach HGL (1828) *Conspectus Regni Vegetabilis* 123.
- Rossow RA (1987) Revisión del género *Mecardonia* (Scrophulariaceae). *Candollea* 42: 431–474.
- Ruiz H, Pavon J (1794) *Flora Peruviana et Chilensis Prodromus*, 95.
- Shaked H, Kashkush K, Ozkan H, Feldman M, Levy AA (2001) Sequence elimination and cytosine methylation are rapid and reproducible responses of the genome to wide hybridization and allopolyploidy in wheat. *The Plant Cell* 13(8): 1749–1759. <https://doi.org/10.2307/3871316>
- Sinha ARP (1987) Report of B chromosome in *Mecardonia procumbens* (Miller) Hassl. *Cytologia* 52: 373–375. <http://doi.org/10.1508/cytologia.52.373>
- Small JK (1903) *Flora of the Southeastern United States*, New York, 1064.
- Schmidel CC (1762) *Icones Plantarum*, Edition Keller 2.
- Soltis DE, Soltis PS, Tate JA (2004) Advances in the study of polyploidy since plant speciation. *New Phytologist* 161(1): 173–191. <https://doi.org/10.1046/j.1469-8137.2003.00948.x>
- Sosa MM, Seijo G (2002) Chromosome Studies in Argentinean Species of *Stemodia* L. (Scrophulariaceae). *Cytologia* 67: 261–266. <http://doi.org/10.1508/cytologia.67.261>
- Sosa MM, Seijo GJ, Fernández A (2009) Chromosome studies in South American species of *Stemodia* (Scrophulariaceae) and their geographical distribution. *Annales Botanici Fennici* 46: 389–396. <http://dx.doi.org/10.5735/085.046.0503>
- Sosa MM, Panseri AF, Fernández A (2011) Karyotype analysis of the southernmost South American species of *Stemodia* (Scrophulariaceae). *Plant Biosystems* 145(2): 472–477. <http://dx.doi.org/10.1080/11263504.2011.566250>
- Sosa M M, Dematteis M (2014) *Stemodia diplohyptoides* (Plantaginaceae, Gratiolae) a new species diploid from the South American. *Phytotaxa* 186(5): 271–278. <http://dx.doi.org/10.11646/phytotaxa.186.5.4>
- Souza VC (1997) Consideraciones sobre la delimitación de *Mecardonia procumbens* (Mill.) Small (Scrophulariaceae). *Acta Botanica Brasílica* 11: 181–189.
- Souza V, Giulietti AM (2009) Levantamento das espécies de Scrophulariaceae *sensu lato* nativas do Brasil. *Pesquisas Botânica* 60: 7–288.
- Subramanian D, Pondmudi R (1987) Cytotaxonomical studies of South Indian *Scrophulariaceae*. *Cytologia* 52: 529–541. <https://doi.org/10.1508/cytologia.52.529>
- Sutton DA (1988) A Revision of the Tribe Antirrhineae, 455–461, f. 122.
- Vargas PJAR, Rosselló JA, Oyama R, Güemes J (2004) Molecular evidence for naturalness of genera in the tribe Antirrhineae (Scrophulariaceae) and three independent evolutionary lineages from the New World and the Old. *Plant Systematics and Evolution* 249: 151–172. <https://doi.org/10.1007/s00606-004-0216-1>
- Vega AJ, Dematteis M (2015) DNA content in species of *Vernonia* and *Vernonanthura* from South America: An approach to systematics and evolution of the Vernonieae (Asteraceae). *Plant Biosystems*, 1–8. <http://dx.doi.org/10.1080/11263504.2015.1115436>
- Wolfe AD, Darwyler SL, Randle CP (2002) A phylogenetic and biogeographic analysis of the Cheloneae (Scrophulariaceae) based on ITS and matK sequence data. *Systematic Botany* 27(1): 138–148. <http://dx.doi.org/10.1043/0363-6445-27.1.138>
- Wong C, Murray BG (2012) Variable changes in genome size associated with different polyploid events in *Plantago* (Plantaginaceae). *Journal of Heredity* 103: 711–719. <https://doi.org/10.1093/jhered/ess049>